

Multi-environment Manifestation of Heterosis for Morphological and Quality Traits in Maize (*Zea mays* L.)

Avinash Kumar¹, N. Kiran¹, Prashant Bisen¹, Amit Dadheech¹, Kaushal Kishor² and Mithilesh Kumar Singh^{3*}

¹Department of Plant Breeding and Genetics, Rajasthan College of Agriculture, Maharana Pratap University of Agriculture and Technology, Udaipur- 313001, India.

²Department of Agronomy, Rajasthan College of Agriculture, Maharana Pratap University of Agriculture and Technology, Udaipur- 313001, India.

³Department of Plant Breeding and Genetics, Dr. Rajendra Prasad Central Agricultural University, Pusa, Bihar -848125, India.

Authors' contributions

This work was carried out in collaboration among all authors. Author AK carried out the experiment and prepared the first hand manuscript. Authors NK, PB and KK carried out collection of reviews and data analysis. Author AD provided guidance during conduction of experiment, while proof reading and correction of manuscript was carried out by author MKS. All authors have read the manuscript thoroughly and approved it.

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ABSTRACT

An investigation was carried out with 10 parents, their 45 hybrids mated in half-diallel fashion and 4 checks in RBD for estimation of heterosis in two environment (*Kharif*, 2014 and *Rabi*, 2014-15) at the Instructional farm, RCA, MPUAT, Udaipur, India. The average productivity of maize in USA is 11.07 MT ha⁻¹ while, in India it is only 3.02 MT ha⁻¹ during 2018-19. We lag far behind USA and are still not able to harness the optimum yield potential in maize that could partly be addressed by evolving hybrid maize cultivar. The present study aims to identify heterotic hybrids that could yield

*Corresponding author: E-mail: mithileshgpb@gmail.com;

stably with better grain quality over a range of environments. Environment wise Analysis of variance revealed that mean squares due to genotypes were significant for all the traits in both the environments. Partitioning of total genotypic variance indicated significant contribution of parents for all the traits. Mean squares due to crosses were significant for all the traits under study except for days to 50% tasseling. Mean squares due to parents vs. crosses were significant for all the traits except for days to 75% brown husk. Bartlett test showed homogeneity of error variance only for one trait *i.e.* grain oil content. The pooled analysis for grain oil content revealed significant differences between the environments. The mean squares due to genotypes including parents, crosses and parents vs. crosses were also significant. Significance of GE interaction for oil content indicates the influence of environments on the expression of this trait. The partitioning of GE interaction into those that by parents \times environment and crosses \times environment interaction were also significant. Two hybrids in E_1 , *viz.*, $P_3 \times P_6$ and $P_2 \times P_4$ has shown significant negative economic heterosis for days to 50% tasseling, days to 50% silking and days to 75% brown husk. None of the hybrids has shown significant negative economic heterosis for these traits in E_2 environment. The hybrid $P_3 \times P_5$ exhibited maximum positive economic heterosis (6.71%) for grain oil content over the best check HQPM-5 on pooled basis. The hybrid $P_5 \times P_7$ (2.78%) in E_1 and $P_5 \times P_{10}$ (2.58%) in E_2 exhibited maximum positive economic heterosis for grain starch content over the best check HQPM-1. The maximum positive economic heterosis for grain protein content was exhibited by hybrid $P_5 \times P_8$ (4.14%) in E_1 and by $P_3 \times P_5$ (2.49%) in E_2 over the best check HQPM-5.

Keywords: Heterosis; environment; quality traits; Zea mays; maize protein; economic heterosis.

1. INTRODUCTION

Maize is the third most widely cultivated cereal crop in the world after Rice and Wheat [1]. It is grown round the year in diverse ecologies and it records highest production and productivity among cereals. It is the primary staple food in many developing countries [2]. It caters to the dietary need of not only the humans but as feed to cattles and as an industrial input. It is believed to be originated in Southern Mexico or Northern Guatemala [3]. Maize grain is becoming increasingly popular in our country due to its huge demand particularly in poultry feed industry. Globally, maize is cultivated on an area of 191.16 mha, with total production of 1122.69 MT and average productivity of 5.87 MTha⁻¹. In India, maize is cultivated on an area of 9.20 mha with a total production of 27.80 MT and average productivity of 3.02 MTha⁻¹[4]. Maize is highly cross pollinated crop and shows handsome magnitude of hybrid vigour for various traits of economic importance. Information on extent of genetic diversity of parents and degree and direction of heterosis proves to be pivotal in developing superior F_1 's. Dominance gene action that culminates into hybrid vigour has direct bearing on the breeding methodology to be adopted for varietal improvement [5]. Maize consumption in World and India has grown up to 1130.60 MT and 27.50 MT, respectively [6] and with the exploitation of heterosis in development of hybrid maize varieties, India has become self

reliant in meeting its domestic demand. Maize improvement program is under technological transition from open pollinated varieties (OPVs) and multi-parent hybrids (MHs) to single cross hybrids. The single cross hybrids exploit heterosis to the maximum extent and it eases maintenance of parental lines as well as hybrid seed production [7]. Nowadays, corn breeders endeavour to capitalize genetic material in order to develop new maize genotypes characterized by high yielding potentiality and better quality. For this, they need thorough knowledge about the type and relative magnitude of genetic variance components and their interaction with environments as well as heterosis for yield and its components. One of the most informative biometrical tool in this concern is diallel analysis which is widely and extensively used [8]. The degree of heterotic effect in F_1 is correlated with genetic diversity of the parental lines. Greater the divergence among the parents, higher is the heterosis and *vice-versa* [9,10]. However, environment can differentially affect the performance of inbred lines and hybrids and distort this relationship [7]. Keeping in view the above facts, the present study was designed with the objective to determine extent and magnitude of heterosis for various morphological and quality traits in maize over the environments that could ultimately be utilized to develop maize hybrids with high yielding potential and better quality.

2. MATERIALS AND METHODS

2.1 Experimental Site and Design

The experimental materials consisted of 10 inbred lines which were crossed in half diallel fashion to generate 45 single cross hybrids, during *Rabi*, 2014 at the Instructional farm, Rajasthan College of Agriculture, Maharana Pratap University of Agriculture and Technology, Udaipur, India. These 45 hybrids, along with 10 parents and 4 standard checks (Table 1) were evaluated in Randomized Block Design with three replications during *Kharif*, 2014 (E_1) and *Rabi*, 2014-15 (E_2). The materials were evaluated in a single row plot of 4 m length, maintaining plant to plant distance of 25 cm. The recommended package of practices of Zone IVA of Rajasthan was adopted to raise a healthy crop stand.

2.2 Data Recording

The data for 8 quantitative traits *viz.*, days to 50% tasseling, days to 50% silking, days to 75% brown husk, plant height, ear height, grain oil content, grain starch content and grain protein content were recorded for both the environments (E_1 and E_2) on 5 randomly selected plants in each entry, except for days to 50% tasseling, days to 50% silking and days to 75% brown husk where, visual observations were recorded on plot basis. The mean data were subjected for statistical analysis.

2.3 Statistical Analysis

The trait mean values were subjected to Analysis of Variance (ANOVA) using the statistical analysis procedures of Panse and

Sukhatme [11]. Average Heterosis [5], Heterobeltiosis [12] and Economic Heterosis [13] were calculated for individual as well as over the environments. The test for homogeneity of error variance for pooled analysis of variance was carried out as procedure suggested by Bartlett [14].

3. RESULTS AND DISCUSSION

Environment wise analysis of variance revealed significant difference among mean of genotypes for all the traits. The total genotypic variance was further partitioned into that assignable to parents, crosses and to parents v_s . crosses. Genotypic variance due to parents was significant for all the traits. Fraction of genotypic variance assignable to the crosses was significant for all the traits under study except for days to 50% tasseling. And proportion of genotypic variance assignable to parents v_s . crosses was significant for all the traits except for days to 75% brown husk. This suggested the presence of heterosis for most of the traits (Table 2). Similar results were observed by Sadaiah [15] and Avinash [16].

Results from Bartlett test indicated that error variance was homogenous only for grain oil content, therefore, pooled analysis was carried out (Table 2). The pooled analysis revealed significant difference among the environments for this trait. The mean sum of squares was significant for all the sources of variation *i.e.* genotypes, crosses and the parents v_s . crosses. Significance of mean sum of squares due to GE interactions for grain oil content indicates the influence of environment on the expression of this trait. The partitioning of GE interaction into

Table 1. List of maize parental inbreds and checks

Sl. No.	Inbred line, Code	Source
1.	EIQ – 105 (P_1)	AICRP* on Maize, Udaipur
2.	EIQ – 106 (P_2)	AICRP* on Maize, Udaipur
3.	EIQ – 107 (P_3)	AICRP* on Maize, Udaipur
4.	EIQ – 108 (P_4)	AICRP* on Maize, Udaipur
5.	EIQ – 109 (P_5)	AICRP* on Maize, Udaipur
6.	EIQ – 110 (P_6)	AICRP* on Maize, Udaipur
7.	EIQ – 111 (P_7)	AICRP* on Maize, Udaipur
8.	EIQ – 112 (P_8)	AICRP* on Maize, Udaipur
9.	EIQ – 113 (P_9)	AICRP* on Maize, Udaipur
10.	EIQ – 114 (P_{10})	AICRP* on Maize, Udaipur
11.	Pratap QPM Hybrid- 1 (Check-1)	AICRP* on Maize, Udaipur
12.	Vivek QPM- 9 (Check-2)	VPKAS [†] , Almora
13.	HQPM- 1 (Check-3)	CCSHAU [‡] , Karnal
14.	HQPM- 5 (Check-4)	CCSHAU [‡] , Karnal

*All India Coordinated Research Project, [†]Vivekanand Parvatiya Krishi Anusandhan Shala and [‡]Choudhary Charan Singh Haryana Agricultural University

that due to parents×environment and crosses×environment were also found to be significant (Table 3).

Among maturity related traits, the exploitable average heterosis ranged from -20.23 ($P_2 \times P_4$) to -2.26 % ($P_1 \times P_3$) in E_1 and from -7.52 ($P_3 \times P_{10}$) to -3.60% ($P_3 \times P_6$) in E_2 for days to 50% tasseling. A total of 36 and 17 hybrids exhibited significant negative average heterosis for this trait in E_1 and E_2 environments, respectively. For days to 50% silking, the exploitable average heterosis ranged from -22.63 ($P_2 \times P_4$) to -2.08% ($P_1 \times P_4$) in E_1 and from -7.65 ($P_3 \times P_{10}$) to -3.46% ($P_4 \times P_9$) in E_2 . A total of 35 and 19 hybrids showed significant negative average heterosis for this trait in E_1 and E_2 environments, respectively. For days to 75% brown husk, the exploitable average heterosis ranged from -13.97 ($P_2 \times P_4$) to -1.72% ($P_3 \times P_8$) in E_1 and 28 hybrids showed significant superiority over the mid parental value. In E_2 , negative significant average heterosis was exhibited only by 3 hybrids viz., $P_1 \times P_6$ (-3.17%), $P_5 \times P_6$ (-2.61%) and $P_8 \times P_9$ (-2.61%) (Table 4).

Among plant type traits, the exploitable average heterosis ranged from -12.41 ($P_2 \times P_4$) to -16.78% ($P_3 \times P_6$) in E_1 while none of the hybrids in E_2 exhibited significant negative average heterosis for plant height. For ear height, 4 hybrids viz., $P_3 \times P_6$ (-32.34%), $P_3 \times P_7$ (-21.03 %), $P_2 \times P_4$ (-20.15 %) and $P_3 \times P_8$ (-16.54%) in E_1 and only one hybrid $P_1 \times P_5$ (-18.88%) in E_2 exhibited significant negative average heterosis for this trait. Two hybrids exhibited significant negative average heterosis for plant height whereas, 4 hybrids exhibited significant negative average heterosis for ear height in E_1 environment. In E_2 environment, none of the hybrid showed significant superiority over average parental value for plant height while one hybrid showed significant superiority over average parental value for ear height (Table 5).

Among quality traits, the exploitable average heterosis for grain oil content ranged from 1.36 ($P_3 \times P_4$) to 102.49% ($P_3 \times P_5$) in E_1 and from 4.95 ($P_4 \times P_6$) to 59.59% ($P_1 \times P_8$) in E_2 . Pooled analysis indicated 35 hybrids to be significantly superior over mid parental value for grain oil content. The number of hybrids exhibiting significant positive average heterosis were 32 and 35 in E_1 and E_2 environment, respectively (Table 7). For grain starch content, the exploitable average heterosis varied from 1.49 ($P_9 \times P_{10}$) to 15.74% ($P_5 \times P_7$) in E_1 and from 0.99 ($P_9 \times P_{10}$) to 16.71% ($P_5 \times P_{10}$) in E_2 environment, respectively. The number of

hybrids exhibiting significant positive average heterosis were 31 and 43 in E_1 and E_2 environment, respectively. For grain protein content, the exploitable average heterosis ranged from 1.18% ($P_1 \times P_4$) to 58.8% ($P_6 \times P_8$) and 40 hybrids showed significant positive average heterosis (Table 6). In E_2 , the exploitable average heterosis ranged from 3.89% ($P_1 \times P_2$) to 53.66% ($P_6 \times P_8$) and 38 hybrids showed significant positive average heterosis. Assuncao [17], Amanullah [18] and Amiruzzaman [19] reported average heterosis for maturity related traits, whereas Ji [20], Muraya [21] and Vieira [22] for plant type traits. Average heterosis for grain oil content and grain starch content was reported by Assuncao [17] and for grain protein content by Yusuf [23].

Perusal of Table 4 showed that 19 hybrids exhibited significant negative heterobeltiosis for days to 50% tasseling, 17 hybrids for days to 50% silking and days to 75% brown husk each in E_1 environment. In E_2 environment, 7 hybrids showed significant negative heterobeltiosis for days to 50% tasseling and 6 hybrids for days to 50% silking. None of the hybrids showed significant negative heterobeltiosis for days to 75% brown husk in E_2 environment. Two hybrids in E_1 and none of the hybrid in E_2 environment exhibited significant negative heterobeltiosis for plant height. Similarly, 2 hybrids in E_1 and none of the hybrid in E_2 environment showed significant negative heterobeltiosis for ear height (Table 5). 25 hybrids showed significant positive heterobeltiosis for grain oil content while 33 hybrids showed significant positive heterobeltiosis for grain starch content in E_1 environment. Similarly, 26 hybrids showed significant positive heterobeltiosis for grain oil content while 37 hybrids showed significant positive heterobeltiosis for grain starch content in E_2 environment. For grain protein content, 32 hybrids in E_1 and 31 hybrids in E_2 showed significant positive heterobeltiosis. The presence of heterobeltiosis indicated that over dominance played an important role in the expression of all these traits. However, its magnitude and number of hybrids which exhibited significant heterobeltiosis were variable. Amanullah [18], Silva [24] and Khanorkar [25] reported heterobeltiosis for maturity and plant type traits.

Among maturity related traits, 2 hybrids in E_1 , viz., $P_3 \times P_6$ and $P_2 \times P_4$ has shown significant negative economic heterosis for days to 50% tasseling, days to 50% silking and days to 75% brown husk against the best check Vivek QPM-9.

Table 2. Mean squares for different traits in maize (*Zea mays* L.)

SN	Traits	Env	Source						Bartlett
			Rep	Genotype	Parent	Crosses	Parents vs crosses	Error	
			[2]	[54]	[9]	[44]	[1]	[108]	
1	Days to 50% tasseling	1	14.8061**	28.2061**	31.8667**	22.7246**	236.446**	0.843098	130.755**
		2	41.2788*	21.7246**	31.8556**	12.169	350.99**	9.55657	
2	Days to 50% silking	1	6.66061**	35.6135**	31.8704**	32.7569**	194.99**	0.808754	132.931**
		2	45.2788**	24.9488**	28.3852**	16.5044**	365.577**	9.3899	
3	Days to 75% brown husk	1	3.07879*	38.6123**	36.6704**	35.7306**	182.885**	0.942985	102.648**
		2	11.4606	17.3912**	36.0926**	13.8832**	3.43468	7.7569	
4	Plant height (cm)	1	177.188*	1211.56**	574.578**	673.232**	30630.8**	56.0768	9.35888**
		2	23.4606	1234.17**	901.07**	378.492**	41882**	30.9915	
5	Ear height (cm)	1	290.036**	275.027**	330.607**	192.818**	3392.01**	27.3018	25.2005**
		2	218.806	450.418**	902.207**	250.415**	5184.44**	73.1085	
12	Grain oil content (%)	1	0.000447602	1.55289**	1.54659**	1.32145**	11.793**	0.00111516	0.3006
		2	0.00381346*	1.63659**	1.29464**	1.45774**	12.5834**	0.00100347	
13	Grain starch content (%)	1	1.12755*	21.3268**	4.01818**	18.6917**	293.05**	0.323773	11.1071**
		2	0.0491102	23.0797**	7.25162**	18.5051**	366.813**	0.16955	
14	Grain protein content (%)	1	0.00373568	3.03108**	2.08455**	2.32621**	42.5642**	0.00426491	47.997**
		2	0.00424793	2.83954**	1.89798**	2.18448**	40.1362**	0.0170065	

*,** Significant at 5 and 1 percent respectively (Model I)

Table 3. Pooled mean squares for grain oil content in maize (*Zea mays* L.)

Traits	Source											Bartlett
	Env	Rep/Env	Genotype	Parents	Crosses	Parents vs crosses	G×E	Parent×Env	Crosses×E	Parents vs Crosses×E	Pool Error	
	[1]	[4]	[54]	[9]	[44]	[1]	[54]	[9]	[44]	[1]	[216]	
Oil content (%)	0.162369**	0.00213053	3.12674**	2.80007**	2.71076**	24.3699**	0.0627409**	0.0411598**	0.0684356**	0.00640804*	0.00105932	0.3006

*,** Significant at 5 and 1 percent respectively (Model I)

Table 4. Extent of heterosis for days to 50% tasseling and days to 50% silking

SN.	Crosses	Env	Days to 50% tasseling			Days to 50% silking			Days to 75% brown husk		
			RH	Hb	EH	RH	Hb	EH	RH	Hb	EH
1	P1 xP2	E1	-3.37**	-1.15		-2.66**	-1.61		0.37		
2	P1xP2	E2	-1.99	-1.34		-0.66	-0.26		0.53		
3	P1xP3	E1	-2.26*			5.15**			4.80**		
4	P1xP3	E2	-1.32			-1.56	-0.26		0.32		
5	P1 xP4	E1	-3.83**	-3.30**		-2.08*	-1.05		-1.97**	-0.72	
6	P1xP4	E2	-1.47	-1.34		-0.66	-0.53		2.93*		
7	P1xP5	E1	-2.79*	-1.14		-3.92**	-3.16**		-3.40**	-2.17*	
8	P1xP5	E2	-0.13			-0.13			-0.11		
9	P1xP6	E1	-2.54*	-0.00		0.00			0.74		
10	P1xP6	E2	-4.17*	-1.60		-3.33	-0.79		-3.17*	-0.87	
11	P1xP7	E1	-1.42			-2.69**	-0.55		-3.66**	-2.59**	
12	P1 xP7	E2	-1.47	-1.34		-0.26			1.53		
13	P1xP8	E1	-7.83**	-2.45		-6.18**			-5.07**	-1.56	
14	P1xP8	E2	-5.00**	-3.48	-0.00	-5.70**	-4.21*	-0.00	-0.64		
15	P1xP9	E1	-7.40**	-7.14**		-6.60**	-6.35**		-5.45**	-5.11**	
16	P1xP9	E2	-2.60			-1.03			1.59		
17	P1xP10	E1	-6.84**	-2.75*		-4.59**	-1.58		-2.46**		
18	P1xP10	E2	-2.45			-2.41			2.33		
19	P2xP3	E1	-4.05**	-3.49**		-3.56**	-1.68		-1.12	-0.75	
20	P2xP3	E2	-3.28	-2.64		-2.98	-2.09		-1.04		
21	P2xP4	E1	-21.23**	-18.97**	-4.08**	-22.63**	-20.97**	-3.92**	-13.97**	-11.57**	-2.87**
22	P2xP4	E2	-1.06	-0.53		-0.00			0.43		
23	P2xP5	E1	-10.29**	-9.77**		-11.87**	-10.22**		-6.72**	-4.10**	
24	P2xP5	E2	-1.33	-0.80		-2.08	-1.57		-0.63	-0.42	
25	P2xP6	E1	-6.05**	-5.78**		-8.65**	-8.15**		-4.51**	-3.79**	
26	P2xP6	E2	-3.75*	-1.85		-4.21*	-2.09		0.73		
27	P2xP7	E1	-3.79**	-2.37		-2.72**	-1.65		-0.37	-0.00	
28	P2xP7	E2	-2.92	-2.40		-3.00	-2.87		2.69*		
29	P2xP8	E1	-0.89			1.14			2.48**		
30	P2 xP8	E2	-3.01	-2.11		-3.48*	-2.35		-0.74	-0.63	
31	P2xP9	E1	-10.36**	-8.05**		-10.40**	-9.68**		-6.64**	-5.60**	

SN.	Crosses	Env	Days to 50% tasseling			Days to 50% silking			Days to 75% brown husk		
			RH	Hb	EH	RH	Hb	EH	RH	Hb	EH
32	P2×P9	E2	-2.46	-0.53		-1.40			0.21		
33	P2×P10	E1	-8.60**	-2.30		-6.19**	-2.15		-0.71		
34	P2 ×P10	E2	-3.33	-0.53		-3.04			-1.36	-0.63	
35	P3×P4	E1	-6.18**	-2.91*		-7.24**	-3.35**		-4.92**	-1.88*	
36	P3×P4	E2	0.13			0.13			1.06		
37	P3×P5	E1	-5.75**	-4.65**		-7.53**	-3.91**		-5.65**	-2.63**	
38	P3×P5	E2	-0.66			-0.64	-0.26		-0.42		
39	P3×P6	E1	-19.42**	-19.19**	-5.44**	-20.11**	-18.99**	-5.23**	-11.70**	-11.36**	-4.10**
40	P3×P6	E2	-3.60*	-2.34		-2.78	-1.54		-0.72	-0.62	
41	P3×P7	E1	1.47			0.83			-0.00		
42	P3×P7	E2	-3.03	-1.87		-4.13*	-3.39		-0.32		
43	P3×P8	E1	-5.07**	-2.45		-4.35**	-0.60		-1.72*		
44	P3×P8	E2	-4.42*	-4.17*		-4.35*	-4.10*		-0.52		
45	P3×P9	E1	-4.79**	-1.74		-1.63			-1.85*	-0.38	
46	P3×P9	E2	-5.40**	-4.17*		-4.05*	-2.82		1.04		
47	P3×P10	E1	-5.95**			-1.84			-0.72		
48	P3×P10	E2	-7.52**	-5.47**		-7.65**	-5.64**		-1.34	-1.04	
49	P4×P5	E1	-6.67**	-4.55**		-9.56**	-9.33**		-8.48**	-8.48**	
50	P4×P5	E2	-1.87	-1.87		-2.34	-1.57		1.07		
51	P4×P6	E1	-7.56**	-4.62**		-8.47**	-5.98**		-7.86**	-4.55**	
52	P4×P6	E2	-4.03*	-1.60		-3.71*	-1.31		1.59		
53	P4×P7	E1	-3.12**			-3.19**	-0.00		-4.16**	-1.85*	
54	P4×P7	E2	0.27			-0.65	-0.26		2.51		
55	P4×P8	E1	-1.44			3.89**			1.11		
56	P4×P8	E2	-4.34*	-2.93		-4.79**	-3.41		1.39		
57	P4×P9	E1	-6.27**	-6.01**		-4.96**	-3.70**		-3.05**	-1.46	
58	P4×P9	E2	-3.77*	-1.33		-3.46*	-1.05		0.42		
59	P4×P10	E1	-7.85**	-4.35**		-5.05**	-3.09**		-2.96**	-1.41	
60	P4×P10	E2	-0.77			-0.76			0.95		
61	P5×P6	E1	-2.58*	-1.73		-5.57**	-3.26**		-4.57**	-1.14	
62	P5×P6	E2	-3.25	-0.80		-2.41	-0.78		-2.61*	-1.48	
63	P5×P7	E1	-4.93**	-2.96*		-6.13**	-3.30**		-5.97**	-3.70**	
64	P5×P7	E2	-1.07	-1.07		-2.20	-1.82		0.54		

SN.	Crosses	Env	Days to 50% tasseling			Days to 50% silking			Days to 75% brown husk		
			RH	Hb	EH	RH	Hb	EH	RH	Hb	EH
65	P5×P8	E1	-2.06			-5.29**			-5.56**	-0.78	
66	P5×P8	E2	-1.18			-2.70	-2.07		1.37		
67	P5×P9	E1	-4.18**	-2.27		-6.28**	-5.29**		-3.77**	-2.19*	
68	P5×P9	E2	-2.47	0.00		-3.18	-1.55		-1.89	-1.06	
69	P5×P10	E1	-4.28**			-5.82**	-3.63**		-6.09**	-4.59**	
70	P5×P10	E2	-3.09			-4.03*	-1.55		0.10		
71	P6×P7	E1	-1.75	-0.59		-1.09	-0.55		0.00		
72	P6×P7	E2	-4.29*	-1.87		-4.59**	-2.60		0.21		
73	P6×P8	E1	-4.76**	-1.84		-4.57**			-1.73*	-0.39	
74	P6×P8	E2	-6.15**	-5.18*		-6.82**	-5.87**		-1.25	-0.42	
75	P6×P9	E1	-5.62**	-2.89*		-2.95**	-1.63		0.00		
76	P6×P9	E2	-5.58**	-5.58**		-5.25**	-5.25**		0.52		
77	P6×P10	E1	-2.96**			-2.59**			1.80*		
78	P6×P10	E2	-5.16**	-4.31*		-4.09*	-3.25		0.41		
79	P7×P8	E1	0.00			2.87**			3.98**		
80	P7×P8	E2	-2.76	-1.33		-3.87*	-2.86		0.21		
81	P7×P9	E1	-0.57			1.89			0.00		
82	P7×P9	E2	-4.81**	-2.40		-5.87**	-3.91		0.75		
83	P7×P10	E1	-5.72**			-2.60**			-5.69**	-1.85*	
84	P7×P10	E2	-1.29			-1.64			3.20*		
85	P8×P9	E1	-1.16			0.85			-0.56		
86	P8×P9	E2	-5.38**	-4.40*		-6.06**	-5.10*		-2.61*	-2.10	
87	P8×P10	E1	-5.82**			-2.72**			-1.28		
88	P8×P10	E2	-4.19*	-2.33		-4.63**	-2.81		-0.21		
89	P9×P10	E1	-6.04**	-2.19		-4.35**	-1.06		-4.59**	-1.46	
90	P9×P10	E2	-0.88			-0.37			0.10		

Table 5. Extent of heterosis for plant height and ear height

SN.	Crosses	Env	Plant height (cm)			Ear height		
			RH	Hb	EH	RH	Hb	EH
1	P1×P2	E1	25.16**		-22.05**	24.41**		-18.13**
2	P1×P2	E2	36.49**		-21.25**	1.74		-24.91**
3	P1×P3	E1	24.08**		-18.76**	9.80		-12.95
4	P1×P3	E2	34.80**		-4.48	-4.74		-15.38*
5	P1×P4	E1	56.49**		-3.29	75.53**		
6	P1×P4	E2	37.80**		-5.85*	9.76		-7.33
7	P1×P5	E1	41.89**		-14.51**	58.74**		-8.29
8	P1×P5	E2	42.73**		-4.29	-18.88**	-9.58	-20.51**
9	P1×P6	E1	9.44		-28.24**	22.09*		-21.24**
10	P1×P6	E2	34.80**		-4.48	-3.82		-7.69
11	P1×P7	E1	27.15**		-11.22**	17.88*		-7.77
12	P1×P7	E2	28.61**		-0.97	-4.84		-9.89
13	P1×P8	E1	58.56**		-2.32	46.08**		
14	P1×P8	E2	42.38**		-7.99**	-4.55		-15.38*
15	P1×P9	E1	42.72**		-13.73**	63.20**		
16	P1×P9	E2	43.96**		-5.85*	6.22		-12.45
17	P1×P10	E1	40.59**		-21.28**	38.89**		-9.33
18	P1×P10	E2	35.65**		-8.77**	-2.89		-7.69
19	P2×P3	E1	31.12**		-8.32*	9.94		-2.59
20	P2×P3	E2	36.99**		-12.28**	29.53*		-29.30**
21	P2×P4	E1	-12.41*	-11.76*	-41.97**	-20.15*	-14.84	-43.52**
22	P2×P4	E2	32.81**		-18.32**	27.01*		-36.26**
23	P2×P5	E1	45.59**		-5.80	48.26**		-0.52
24	P2×P5	E2	55.66**		-6.24*	37.93**		-12.09
25	P2×P6	E1	26.52**		-11.41**	34.04**		-1.04
26	P2×P6	E2	55.25**		-0.58	64.39**		
27	P2×P7	E1	15.89**		-13.93**	9.47		-4.15
28	P2×P7	E2	22.50**		-14.04**	42.42**		-13.92
29	P2×P8	E1	39.09**		-8.12*	25.84**		
30	P2×P8	E2	58.52**		-8.38**	38.72**		-24.54**
31	P2×P9	E1	30.55**		-15.28**	30.77**		-3.11

SN.	Crosses	Env	Plant height (cm)			Ear height		
			RH	Hb	EH	RH	Hb	EH
32	P2×P9	E2	50.42**		-11.89**	66.54**		-19.78*
33	P2×P10	E1	41.44**		-14.51**	30.56**		-2.59
34	P2×P10	E2	32.58**		-19.88**	35.54**		-17.58*
35	P3×P4	E1	33.70**		-7.16	9.54		-7.77
36	P3×P4	E2	25.07**		-6.63*	32.58**		-13.55
37	P3×P5	E1	37.61**		-6.58	22.83**		-1.04
38	P3×P5	E2	22.97**		-9.75**	20.47*		-5.13
39	P3×P6	E1	-16.78**	-16.67**	-39.07**	-32.34**	-18.57*	-40.93**
40	P3×P6	E2	13.38**		-12.48**	28.40**		-1.47
41	P3×P7	E1	9.61*		-15.09**	-21.03**	-20.21**	-20.21**
42	P3×P7	E2	24.21**			28.16**		-3.30
43	P3×P8	E1	34.92**		-6.58	-16.54**	-13.59	-17.62**
44	P3×P8	E2	41.48**			42.48**		-1.10
45	P3×P9	E1	41.19**		-3.87	17.75**		
46	P3×P9	E2	45.92**			49.57**		-5.49
47	P3×P10	E1	39.51**		-11.22**	1.18		-10.88
48	P3×P10	E2	30.33**		-4.09	18.84*		-9.89
49	P4×P5	E1	37.05**		-11.99**	46.28**		-8.29
50	P4×P5	E2	34.25**		-4.87	24.14**		-7.69
51	P4×P6	E1	34.91**		-6.19	50.75**		
52	P4×P6	E2	26.11**		-5.85*	16.46		-15.75*
53	P4×P7	E1	24.51**		-8.12*	32.71**		
54	P4×P7	E2	17.01**		-5.46*	4.64		-25.64**
55	P4×P8	E1	36.58**		-10.44**	15.38*		-6.74
56	P4×P8	E2	28.77**		-11.89**	21.69*		-20.88**
57	P4×P9	E1	35.74**		-12.57**	22.68**		-14.51*
58	P4×P9	E2	48.17**			57.01**		-7.69
59	P4×P10	E1	25.81**		-24.56**	17.34*		-17.62**
60	P4×P10	E2	29.22**		-8.19**	23.59*		-11.72
61	P5×P6	E1	26.88**		-13.73**	53.54**		
62	P5×P6	E2	38.91**			16.42*		0.00
63	P5×P7	E1	35.21**		-2.32	40.72**		
64	P5×P7	E2	27.21**			11.26		-5.86

SN.	Crosses	Env	Plant height (cm)			Ear height		
			RH	Hb	EH	RH	Hb	EH
65	P5×P8	E1	46.53**		-6.19	41.61**		
66	P5×P8	E2	30.62**		-12.28**	27.27**		0.00
67	P5×P9	E1	48.31**		-6.77	72.55**		
68	P5×P9	E2	57.25**			55.44**		
69	P5×P10	E1	52.32**		-11.03**	55.64**		
70	P5×P10	E2	44.69**			-3.88	-0.45	-18.32*
71	P6×P7	E1	5.49		-18.18**	8.11		-6.74
72	P6×P7	E2	20.94**			25.94**		
73	P6×P8	E1	43.93**		-0.19	32.72**		
74	P6×P8	E2	40.66**		-0.19	32.06**		
75	P6×P9	E1	39.01**		-5.22	45.91**		
76	P6×P9	E2	38.04**		-0.97	48.96**		
77	P6×P10	E1	23.52**		-21.28**	31.45**		-3.63
78	P6×P10	E2	34.57**		-0.97	9.49		-9.16
79	P7×P8	E1	20.11**		-11.61**	0.27		-2.07
80	P7×P8	E2	28.45**		-0.97	18.73*		-10.62
81	P7×P9	E1	23.63**		-10.44**	27.54**		
82	P7×P9	E2	31.66**			51.72**		
83	P7×P10	E1	29.45**		-11.99**	13.69*		-1.04
84	P7×P10	E2	15.89**		-7.60**	11.66		-8.79
85	P8×P9	E1	19.28**		-23.40**	4.00		-12.44
86	P8×P9	E2	30.06**		-14.81**	43.02**		-9.89
87	P8×P10	E1	42.39**		-14.89**	18.04**		-0.00
88	P8×P10	E2	34.59**		-9.36**	35.11**		
89	P9×P10	E1	45.21**		-14.89**	28.87**		-5.18
90	P9×P10	E2	35.62**		-7.60**	16.62		-19.05*

Table 6. Extent of heterosis for grain starch content (%) and grain protein content (%)

SN.	Crosses	Env	Grain starch content (%)			Grain protein content (%)		
			RH	Hb	EH	RH	Hb	EH
1	P1×P2	E1	6.03**	4.79**		6.75**	1.08	
2	P1×P2	E2	6.08**	4.25**		3.89**		
3	P1×P3	E1	2.76**	0.12		0.24		
4	P1×P3	E2	4.25**	0.29		-0.06		
5	P1×P4	E1	1.72*			1.18*	0.53	
6	P1×P4	E2	2.02**			1.19	0.86	
7	P1×P5	E1	10.21**	9.69**		4.79**	4.29**	
8	P1×P5	E2	13.80**	13.48**		0.70	0.45	
9	P1×P6	E1	6.21**	4.23**		25.83**	10.62**	
10	P1×P6	E2	9.71**	6.56**		18.92**	4.98**	
11	P1×P7	E1	3.92**	3.08**		-6.92**		
12	P1×P7	E2	3.54**	1.41*		-5.08**		
13	P1×P8	E1	13.82**	11.48**	1.93**	35.97**	29.54**	2.03**
14	P1×P8	E2	12.33**	8.65**		32.69**	26.65**	0.79
15	P1×P9	E1	1.95**	0.82		10.93**		
16	P1×P9	E2	1.39**	0.03		10.94**		
17	P1×P10	E1	5.13**	2.29**		8.17**	8.17**	
18	P1×P10	E2	8.55**	5.62**		11.31**	7.62**	
19	P2×P3	E1	5.24**	3.73**		8.65**	0.28	
20	P2×P3	E2	4.37**	2.13**		8.17**	0.87	
21	P2×P4	E1	7.28**	6.18**		16.17**	9.34**	
22	P2×P4	E2	6.45**	5.88**		11.94**	6.26**	
23	P2×P5	E1	9.03**	8.26**		-8.23**		
24	P2×P5	E2	12.23**	10.61**		-10.31**		
25	P2×P6	E1	-2.60**			23.28**	13.94**	
26	P2×P6	E2	-1.50**			23.35**	13.89**	
27	P2×P7	E1	13.31**	12.89**	1.34	21.52**	20.42**	
28	P2×P7	E2	10.66**	10.28**		17.54**	15.03**	
29	P2×P8	E1	2.04**	1.11		20.52**	19.74**	
30	P2×P8	E2	3.10**	1.44**		15.88**	15.59**	
31	P2×P9	E1	8.46**	8.40**		25.88**	16.78**	

SN.	Crosses	Env	Grain starch content (%)			Grain protein content (%)		
			RH	Hb	EH	RH	Hb	EH
32	P2×P9	E2	8.42**	7.99**		28.84**	19.22**	
33	P2×P10	E1	3.92**	2.29**		16.08**	9.92**	
34	P2×P10	E2	4.77**	3.71**		18.80**	16.94**	
35	P3×P4	E1	5.00**	4.57**		7.83**	5.60**	
36	P3×P4	E2	6.41**	4.68**		5.96**	3.98**	
37	P3×P5	E1	7.60**	5.32**		26.28**	22.32**	1.81**
38	P3×P5	E2	9.27**	5.42**		25.61**	23.16**	2.49*
39	P3×P6	E1	8.62**	7.83**		22.41**	5.13**	
40	P3×P6	E2	6.94**	5.90**		23.33**	6.81**	
41	P3×P7	E1	-0.31			9.00**		
42	P3×P7	E2	-1.68**			7.22**		
43	P3×P8	E1	1.85**	1.31		3.46**		
44	P3×P8	E2	1.82**	1.25*		6.49**		
45	P3×P9	E1	9.08**	7.45**		44.18**	24.25**	3.41**
46	P3×P9	E2	8.14**	5.41**		40.14**	21.62**	1.21
47	P3×P10	E1	3.20**	3.06**		-8.51**		
48	P3×P10	E2	4.68**	3.46**		-9.57**		
49	P4×P5	E1	7.15**	5.31**		6.41**	5.22**	
50	P4×P5	E2	7.98**	5.86**		5.41**	5.32**	
51	P4×P6	E1	2.82**	2.49**		16.58**	1.93**	
52	P4×P6	E2	4.61**	3.91**		19.22**	4.95**	
53	P4×P7	E1	5.41**	3.94**		16.07**	8.31**	
54	P4× P7	E2	6.89**	6.68**		8.23**	0.65	
55	P4×P8	E1	7.32**	7.19**		-7.17**		
56	P4×P8	E2	6.72**	5.57**		-7.20**		
57	P4×P9	E1	0.47			14.91**	0.82	
58	P4×P9	E2	3.30**	2.35**		13.96**	0.53	
59	P4×P10	E1	-2.15**			6.52**	5.84**	
60	P4×P10	E2	0.99*	0.51		8.21**	4.30**	
61	P5×P6	E1	9.23**	7.69**		24.95**	10.31**	
62	P5×P6	E2	11.54**	8.64**		21.33**	6.89**	
63	P5×P7	E1	15.74**	15.35**	2.78**	30.54**	23.10**	
64	P5×P7	E2	13.91**	11.88**		29.99**	20.98**	

SN.	Crosses	Env	Grain starch content (%)			Grain protein content (%)		
			RH	Hb	EH	RH	Hb	EH
65	P5×P8	E1	12.79**	10.99**	1.48*	39.74**	33.74**	4.33**
66	P5×P8	E2	14.65**	11.21**	1.41**	33.25**	26.89**	1.47
67	P5×P9	E1	4.93**	4.25**		26.35**	11.95**	
68	P5×P9	E2	5.88**	4.76**		30.69**	15.37**	
69	P5×P10	E1	13.19**	10.65**	2.53**	12.50**	11.96**	
70	P5×P10	E2	16.71**	13.88**	2.58**	14.21**	10.16**	
71	P6×P7	E1	7.70**	6.54**		35.97**	26.73**	
72	P6×P7	E2	6.25**	5.35**		31.43**	23.83**	
73	P6×P8	E1	12.34**	12.11**	2.51**	58.58**	45.70**	3.91**
74	P6×P8	E2	12.90**	12.43**	2.52**	53.66**	41.55**	2.39*
75	P6×P9	E1	4.70**	3.89**		42.49**	41.91**	
76	P6×P9	E2	6.93**	5.24**		41.24**	40.90**	
77	P6×P10	E1	0.05			25.40**	10.25**	
78	P6×P10	E2	1.76**	1.56**		28.65**	17.07**	
79	P7×P8	E1	3.93**	2.61**		36.62**	34.51**	
80	P7×P8	E2	5.16**	3.83**		38.04**	34.75**	
81	P7×P9	E1	5.22**	4.89**		34.33**	25.69**	
82	P7×P9	E2	3.95**	3.18**		40.10**	32.30**	
83	P7×P10	E1	1.29			28.50**	20.62**	
84	P7×P10	E2	3.37**	2.68**		30.24**	25.50**	
85	P8×P9	E1	3.11**	2.11**		46.01**	34.65**	
86	P8×P9	E2	2.34**	0.30		43.31**	32.31**	
87	P8×P10	E1	3.15**	2.47**		4.13**		
88	P8×P10	E2	1.91**	1.29*		12.07**	10.59**	
89	P9×P10	E1	1.49*			17.92**	4.04**	
90	P9×P10	E2	2.02**	0.60		19.32**	8.82**	

Table 7. Extent of heterosis for grain oil content (%)

SN.	Crosses	Env	Grain oil content (%)		
			RH	Hb	EH
1	P1×P2	E1	16.47**	15.59**	
2	P1×P2	E2	10.74**	10.08**	
3	P1×P2	Pool	13.58**	12.81**	
4	P1×P3	E1	32.32**	30.22**	
5	P1×P3	E2	33.82**	29.70**	
6	P1×P3	Pool	33.07**	29.95**	
7	P1×P4	E1	12.57**		
8	P1×P4	E2	11.43**		
9	P1×P4	Pool	12.00**		
10	P1×P5	E1	48.99**	43.78**	
11	P1×P5	E2	43.88**	39.60**	
12	P1×P5	Pool	46.41**	41.67**	
13	P1×P6	E1	-17.68**		
14	P1×P6	E2	-13.12**		
15	P1×P6	Pool	-15.40**		
16	P1×P7	E1	16.72**		
17	P1×P7	E2	25.23**	6.02**	
18	P1×P7	Pool	20.92**	0.96*	
19	P1×P8	E1	64.11**	51.12**	0.98*
20	P1×P8	E2	59.59**	43.91**	1.84**
21	P1×P8	Pool	61.80**	47.40**	1.41**
22	P1×P9	E1	43.98**	33.98**	
23	P1×P9	E2	43.34**	37.25**	
24	P1×P9	Pool	43.66**	35.58**	
25	P1×P10	E1	27.96**	21.84**	
26	P1×P10	E2	26.44**	17.27**	
27	P1×P10	Pool	27.19**	19.47**	
28	P2×P3	E1	23.21**	20.34**	
29	P2×P3	E2	26.88**	22.26**	
30	P2×P3	Pool	25.05**	21.31**	
31	P2×P4	E1	38.22**	23.22**	

SN.	Crosses	Env	Grain oil content (%)		
			RH	Hb	EH
32	P2×P4	E2	39.20**	23.91**	
33	P2×P4	Pool	38.71**	23.57**	
34	P2×P5	E1	54.25**	47.77**	
35	P2×P5	E2	59.54**	53.89**	
36	P2×P5	Pool	56.93**	50.85**	
37	P2×P6	E1	-12.20**		
38	P2×P6	E2	-7.54**		
39	P2×P6	Pool	-9.87**		
40	P2×P7	E1	15.05**		
41	P2×P7	E2	13.83**		
42	P2×P7	Pool	14.45**		
43	P2×P8	E1	31.81**	22.23**	
44	P2×P8	E2	36.97**	24.19**	
45	P2×P8	Pool	34.44**	23.24**	
46	P2×P9	E1	22.76**	15.04**	
47	P2×P9	E2	19.87**	15.43**	
48	P2×P9	Pool	21.32**	15.24**	
49	P2×P10	E1	30.33**	25.00**	
50	P2×P10	E2	17.78**	9.84**	
51	P2×P10	Pool	23.91**	17.14**	
52	P3×P4	E1	1.36*		
53	P3×P4	E2	8.72**		
54	P3×P4	Pool	5.05**		
55	P3×P5	E1	102.49**	98.51**	8.09**
56	P3×P5	E2	97.20**	96.99**	5.34**
57	P3×P5	Pool	99.84**	97.97**	6.71**
58	P3×P6	E1	-2.72**		
59	P3×P6	E2	5.47**		
60	P3×P6	Pool	1.35**		
61	P3×P7	E1	-2.00**		
62	P3×P7	E2	-3.10**		
63	P3×P7	Pool	-2.54**		
64	P3×P8	E1	-8.96**		

SN.	Crosses	Env	Grain oil content (%)		
			RH	Hb	EH
65	P3×P8	E2	-5.41**		
66	P3×P8	Pool	-7.16**		
67	P3×P9	E1	22.59**	12.39**	
68	P3×P9	E2	27.30**	18.30**	
69	P3×P9	Pool	24.91**	15.28**	
70	P3×P10	E1	41.72**	32.90**	
71	P3×P10	E2	37.81**	24.18**	
72	P3×P10	Pool	39.73**	28.38**	
73	P4×P5	E1	26.07**	8.24**	
74	P4×P5	E2	21.26**	4.60**	
75	P4×P5	Pool	23.64**	6.40**	
76	P4×P6	E1	-1.24*		
77	P4×P6	E2	4.95**		
78	P4×P6	Pool	1.86**		
79	P4×P7	E1	-2.50**		
80	P4×P7	E2	-7.45**		
81	P4×P7	Pool	-4.95**		
82	P4×P8	E1	37.30**	31.54**	
83	P4×P8	E2	40.38**	37.54**	1.44**
84	P4×P8	Pool	38.87**	34.56**	
85	P4×P9	E1	17.64**	11.49**	
86	P4×P9	E2	26.37**	16.43**	
87	P4×P9	Pool	21.98**	13.98**	
88	P4×P10	E1	-5.39**		
89	P4×P10	E2	-15.32**		
90	P4×P10	Pool	-10.46**		
91	P5×P6	E1	6.73**		
92	P5×P6	E2	11.23**		
93	P5×P6	Pool	8.98**		
94	P5×P7	E1	17.36**		
95	P5×P7	E2	13.27**		
96	P5×P7	Pool	15.33**		
97	P5×P8	E1	61.09**	43.60**	

SN.	Crosses	Env	Grain oil content (%)		
			RH	Hb	EH
98	P5×P8	E2	44.34**	26.70**	
99	P5×P8	Pool	52.51**	34.88**	
100	P5×P9	E1	28.16**	15.40**	
101	P5×P9	E2	37.61**	28.00**	
102	P5×P9	Pool	32.86**	21.56**	
103	P5×P10	E1	41.85**	30.58**	
104	P5×P10	E2	42.56**	28.58**	
105	P5×P10	Pool	42.21**	29.54**	
106	P6×P7	E1	-27.11**		
107	P6×P7	E2	-31.54**		
108	P6×P7	Pool	-29.29**		
109	P6×P8	E1	39.18**	23.66**	6.36**
110	P6×P8	E2	38.13**	27.19**	6.95**
111	P6×P8	Pool	38.65**	25.41**	6.66**
112	P6×P9	E1	-4.13**		
113	P6×P9	E2	-4.56**		
114	P6×P9	Pool	-4.34**		
115	P6×P10	E1	-0.78		
116	P6×P10	E2	0.53		
117	P6×P10	Pool	-0.12		
118	P7×P8	E1	9.82**		
119	P7×P8	E2	19.32**	11.13**	
120	P7×P8	Pool	14.57**	3.96**	
121	P7×P9	E1	-4.98**		
122	P7×P9	E2	-6.77**		
123	P7×P9	Pool	-5.85**		
124	P7×P10	E1	-0.12		
125	P7×P10	E2	5.34**		
126	P7×P10	Pool	2.61**		
127	P8×P9	E1	4.55**	3.37**	
128	P8×P9	E2	5.70**		
129	P8×P9	Pool	5.13**	1.26**	
130	P8×P10	E1	34.05**	29.41**	

SN.	Crosses	Env	Grain oil content (%)		
			RH	Hb	EH
131	P8×P10	E2	27.62**	23.78**	
132	P8×P10	Pool	30.72**	26.51**	
133	P9×P10	E1	27.83**	24.78**	
134	P9×P10	E2	23.30**	19.26**	
135	P9×P10	Pool	25.55**	24.93**	

Table 8. Crosses exhibiting high economic heterosis in both the environments, E₁ and E₂

Traits	Crosses
Grain oil content	P ₁ ×P ₈ , P ₃ ×P ₅ , P ₆ ×P ₈
Grain starch content	P ₅ ×P ₈ , P ₅ ×P ₁₀ , P ₆ ×P ₈
Grain protein content	P ₃ ×P ₅ , P ₆ ×P ₈

None of the hybrids has shown significant negative economic heterosis for these traits in E_2 environment. Therefore, these 2 hybrids can be recommended for their large scale multi-location testing in *Kharif* season (E_1) for the development of early maturing hybrids (Table 4).

For plant height, the exploitable economic heterosis ranged from -41.97 ($P_2 \times P_4$) to -8.12% ($P_2 \times P_8$ and $P_4 \times P_7$) and 32 hybrids showed significant negative economic heterosis over the best check (HQPM-5) in E_1 environment. Whereas, in E_2 environment, the exploitable economic heterosis ranged from -21.25 ($P_1 \times P_2$) to -5.46% ($P_4 \times P_7$) and 24 hybrids exhibited significant negative economic heterosis over the best check Pratap QPM Hybrid-1. The hybrid $P_2 \times P_4$ (-41.97%) manifested maximum negative economic heterosis in E_1 environment, whereas, the hybrid $P_1 \times P_2$ (-21.25%) manifested maximum negative economic heterosis in E_2 environment.

Eight hybrids with magnitude ranging from -43.52 ($P_2 \times P_4$) to -17.62% ($P_4 \times P_{10}$) and 14 hybrids with magnitude ranging from -36.26 ($P_2 \times P_4$) to -15.38% ($P_1 \times P_3$ and $P_1 \times P_8$) exhibited significant negative economic heterosis for ear height over the best check Vivek QPM-9 in E_1 and Pratap QPM Hybrid-1 in E_2 , respectively. The hybrid $P_2 \times P_4$ exhibited maximum negative economic heterosis in both environments, the E_1 (-43.52%) and the E_2 (-36.26%) (Table 5).

Among quality traits, only 3 hybrids showed significant positive economic heterosis for grain oil content. The hybrid $P_3 \times P_5$ exhibited maximum positive economic heterosis (6.71%) for grain oil content over the best check HQPM-5 on pooled basis. Five hybrids in E_1 and three hybrids in E_2 showed significant positive economic heterosis for grain starch content. The hybrid $P_5 \times P_7$ (2.78%) in E_1 and $P_5 \times P_{10}$ (2.58%) in E_2 exhibited maximum positive economic heterosis for grain starch content over the best check HQPM-1. In case of grain protein content, 5 hybrids in E_1 and 2 hybrids in E_2 exhibited significant positive economic heterosis for this trait. The maximum positive economic heterosis for this trait was exhibited by hybrid $P_5 \times P_8$ (4.14%) in E_1 and by $P_3 \times P_5$ (2.49%) in E_2 over the best check HQPM-5. Abuali [26], Singh [27], Chahar [28], Lahane [29] and Bisen [30] reported economic heterosis to the similar tunes for quality traits in maize.

4. CONCLUSION

It is inferred from the above findings that heterotic effects were evident for maturity, plant type and quality traits in maize hybrids indicating

genetic diversity existing among the parents involved in these crosses. For grain oil content, the same three hybrids ($P_1 \times P_8$, $P_3 \times P_5$, $P_6 \times P_8$) have shown significant positive economic heterosis in pooled environment. Further, for feasible exploitation of above single cross hybrids in the field for quality traits, it is wise to recommend their large scale multi location testing to have an idea about their stability in performance over environment to be ultimately used as popular hybrids.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Poehlman JM. Breeding Field Crops. 5th Edn. The AVI publish. Co. Inc. Westport, Connecticut; 2006.
- Morris DL. Quantitative determination of carbohydrate with Derwood's anthrone reagent. Science. 1948;107:254-255.
- Weatherwax P. History and origin of corn. In G.F. Spragne (Ed.). Corn and corn improvement, Academic Press. New York. 1955;1-16.
- Foreign Agricultural Service/USDA, Office of Global Analysis; 2019.
- Shull GH. What is heterosis. Genetics. 1909;33:439-446.
- USDA FAS Grain: World Markets and Trade; 2019.
- Kumar P, Singh NK, Jha SK. Multi-environment evaluation for determining grain yield, combining ability, heterosis and their inter-relationships in maize. SABRAO J. Breed. Genet., 2015; 47(4):366-374.
- Abdel-Moneam MA, Sultan MS, Sadek SE, Shalof MS. Combining abilities for yield and yield components in diallel crosses of six new yellow maize inbred lines. International Journal of Plant Breeding and Genetics, 2015;9:86-94.
- Prasad SK, Singh TP. Heterosis in relation to genetic divergence in maize (*Zea mays* L.). Euphytica. 1986;35:919-924.
- Duvick DN. Heterosis: feeding people and protecting resources. In: Coors JG, Pandey S. (eds.) The Genetics and Exploitation of Heterosis In Crops. ASSA/CSSA/SSA, Madison WI. 1999;19-29.
- Panse VG, Sukhatme PV. Statistical methods for agriculture workers. ICAR, New Delhi. 1985;14-33.

12. Fonseca S, Patterson FL. Hybrid vigour in a seven parent diallel cross in common winter wheat (*Triticum aestivum* L.). *Crop Science*. 1968;8:85-88.
13. Meredith WR, Bridge RR. Heterosis and gene action in cotton (*G. hirsutum* L.). *Crop Science*. 1972;12:304-310.
14. Bartlett MS. Properties of sufficiency and statistical tests. *Proc. Roy. Soc. London A*. 1937;160:268-282.
15. Sadaiah K, Reddy V, Narsimha and Kumar SS. Heterosis and combining ability studies for sugar content in sweet corn. (*Zea mays saccharata* L.). *International Journal of Scientific and Research Publications*. 2013;3(3):2250-3153.
16. Avinash HA, Jaiwar SS, Girase VK, Rawool SA, Khanorkar SM. Assessment of heterosis and combining ability for biochemical components in crosses among high quality protein maize (*Zea mays* L.). *Journal of Soils and Crops*. 2013;23(1):176-184.
17. Assuncao A, Brasil EM, Oliviera JP, de Reis AJ, dos S, Periera AF, Bueno LG, Ramos MR. Heterosis performance in industrial and yield components of sweet corn. *Crop Breeding and Applied Biotechnology*. 2010;10(3):183-190.
18. Amanullah SJ, Muhammad M, Muhammad AK. Heterosis studies in diallel crosses of maize. *Sarhad Journal of Agriculture*. 2011;27(2):207-211.
19. Amiruzzaman M, Islam MA, Lutful H, Monjurul K, Rohman MM. Heterosis and combining ability in a diallel among elite inbred lines of maize (*Zea mays* L.). *Emirates Journal of Food and Agriculture*. 2013;25(2):132-137.
20. Ji, HC, Cho, JW, Yamakawa T. Diallel analysis of plant and ear heights in tropical maize (*Zea mays* L.). *Journal of the Faculty of Agriculture, Kyushu University*. 2006;51:233-238.
21. Muraya MM, Ndirangu CM, Omolo EO. Heterosis and combining ability in diallel crosses involving maize (*Zea mays* L.) S₁ lines. *Australian Journal of Experimental Agriculture*. 2006;46:387-394.
22. Vieira RA, Souza-neto IL, Bignotto LS, Cruz CD, Amral AT, Scapim CA. Heterotic parametrization for economically important traits in popcorn. *Acta Scientiarum, Agronomy*. 2009;31:411-419.
23. Yusuf M, Ado SG, Isthikyak MF. Heterosis in single crosses of quality protein maize inbred lines. *African crop conference proceeding*, 2009;9:439-445.
24. Silva VQR, Amaral JAT, Gonçalves LSA, Freitas JSP, Candido LS, Vittorazzi C, Moterle LM, Vieira RA, Scapim CA. Heterotic parameterizations of crosses between tropical and temperate lines of popcorn. *Acta Scientiarum, Agronomy, Maringa*. 2011;33(2):243-249.
25. Khanorkar SM, Avinash HA, Jaiwar SS, Girase VK. Heterosis in quality protein maize (*Zea mays* L.). *Maize Journal*. 2012;1(1):30-34.
26. Abuali AI, Abdelmulla AA, Khalafalla MM, Idris AE, Osman AM. Combining ability and heterosis for yield and yield components in maize (*Zea mays* L.). *Australian Journal of Basic and Applied Sciences*. 2012;6(10):36-41.
27. Singh A, Shahi JP, Langade DM. Combining ability studies for yield and its related traits in inbred lines of maize (*Zea mays* L.). *Molecular Plant Breeding*. 2013;4(22):177-188.
28. Chahar S, Vyas M, Ranwah BR, Vadodariya G. Heterosis and combining ability analysis for yield and its contributing traits in early maturing maize (*Zea mays* L.) genotypes. *Trends in Biosciences*. 2014;7(14):1774-1777.
29. Lahane GR, Chauhan RM, Patel JM. Combining ability and heterosis studies for yield and quality traits in quality protein maize. *Journal of Agrisearch*. 2014;1(3):135-138.
30. Bisen P, Dadheech A, Namrata, Nagar O, Meena RK. Exploitation of heterosis in single cross hybrids of quality protein maize (*Zea mays* L.) for yield and quality traits. *International Journal of Bio-resource and Stress Management*. 2017;8(1):012019.

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