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Genomic Innovations in Wheat (*Triticum aestivum* L.): A Comprehensive Review of Recent Developments and Future Directions

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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Review Article

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ABSTRACT

Genomic innovations have revolutionized the field of wheat breeding by providing advanced tools to enhance its biofortification and overall productivity. This comprehensive review synthesizes recent developments in wheat genomics, focusing on the integration of various approaches such as genome-wide association studies (GWAS), marker-assisted selection (MAS), and quantitative trait locus (QTL) mapping. These genomic techniques have significantly advanced our understanding of

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the genetic basis underlying key traits, facilitating the development of wheat varieties with improved nutritional profiles and agronomic characteristics. Association mapping, leveraging natural variation within wheat populations, has identified numerous genetic loci associated with important traits, including grain quality and resistance to abiotic and biotic stresses. Marker-assisted selection (MAS) has been instrumental in accelerating the breeding process by enabling the selection of desirable traits based on molecular markers linked to these traits. Additionally, QTL mapping has provided valuable insights into the genetic architecture of complex traits, guiding breeders in the development of high-vielding and nutritionally enhanced wheat varieties. The convergence of these genomic approaches, including GWAS, MAS, and QTL mapping, has ushered in a new era of precision breeding. This review highlights the synergy between these methodologies, illustrating their combined impact on improving wheat biofortification. Future directions in wheat genomics will likely focus on integrating these approaches with emerging technologies, such as CRISPR-based genome editing and advanced phenotyping techniques, to address the challenges of global food security and climate change. The continuous advancement of these genomic tools promises to further enhance the resilience and nutritional quality of wheat, ensuring its pivotal role in global agriculture.

Keywords: Biofortification; productivity; GWAS; MAS; QTL; association mapping; CRISPR.

1. INTRODUCTION

Common wheat (*Triticum aestivum* L., 2n = 6x =42. AABBDD) is a crucial global crop, providing more than 30% of the world's total calorie intake [1-2]. It is a key component in a variety of cerealbased processed foods, including bread, cookies, and noodles. Wheat is cultivated, consumed, and traded worldwide, ranking third in cereal production. It contains significant amounts of carbohydrates (78.1%), protein (14.7%), fat (2.1%), and minerals (2.1%) [3]. Micronutrient deficiencies are a primary cause of global profoundly malnutrition and affect diets. especially in developing regions. Hidden hunger, prevalent less developed countries. in significantly impacts health. The importance of addressing mineral micronutrient deficiencies, particularly iron and zinc, is critical. Iron deficiency anemia, affecting many women and children of reproductive age, is a notable issue [4]. In populations where cereal-based foods like wheat are dietary staples, the severity of iron and zinc deficiencies is heightened due to the loss of these micronutrients during grain processing, leading to "hidden hunger" [5].

Despite considerable increases in production since the 1960s 'Green Revolution' and the use of marker-assisted molecular breeding [6], wheat faces unprecedented challenges from global climate change, rapid population growth, and water shortages in arid and semi-arid regions [7-8]. Additionally, the excessive use of fertilizers and pesticides exacerbates environmental degradation and pollution. The hexaploid nature and gene redundancy of wheat further

complicate genetic selection, often extending the process or making it difficult due to gene linkage or drag [9]. To ensure global food and ecosystem security, it is crucial to enhance the resilience of wheat production while minimizing environmental impacts through the adoption of advanced technologies. Wheat is a major staple crop worldwide, occupying about 17% of global crop acreage [10]. It is essential for feeding nearly half of the world's population and provides roughly one-fifth of the total food calories and protein in human diets [11]. Although wheat production has seen steady growth over the past forty years, recent trends indicate a decline, with current global wheat stocks reaching their lowest levels since 1948/49.

Genome-editing technologies have revolutionized plant research, offering substantial potential for crop improvement. Among these Clustered technologies, Regularly the Interspaced Short Palindromic Repeat (CRISPR)-CRISPR-associated protein (CRISPR-Cas) system is particularly notable for its versatility, simplicity, and cost-effectiveness in making precise DNA modifications [12]. This system enables targeted mutagenesis, including gene knockout, single-base substitutions, and gene or allele replacements in vivo. Recently, CRISPR/Cas has become a leading tool in genome editing, driving progress in the field. Its applications are especially relevant for advancing plant biology and crop improvement in the face of global climate change and pressing agricultural [14], and [13]. environmental ecological challenges [15]. Leveraging CRISPR/Cas technology presents a promising path toward

addressing these issues, paving the way for precision breeding and sustainable agriculture [16].

Various genomic methods, such as quantitative trait loci (QTL) mapping, marker-assisted selection (MAS), and genomic selection, have been extensively used to improve wheat biofortification [17]. While numerous techniques are available for QTL mapping in experimental crosses, identifying the molecular basis of QTLs remains challenging. This is true even in wellstudied model plants like Arabidopsis and rice, due to the difficulty of precisely pinpointing QTLs to single genes [18]. Factors such as experimental design, the type of plant population studied, and the level of polymorphism between parental genomes also affect QTL predictions. Statistical approaches for identifying QTLs require a large number of molecular markers and high-resolution aenetic maps [19]. This methodology is fundamental to genomics, aimed at unraveling complex phenotypes. Several QTL mapping studies have successfully pinpointed a variety of stable and reliable QTLs, providing valuable insights into the genetic basis of biofortification traits. These discoveries enhance our understanding of wheat genetics and offer promising avenues for future breeding programs focused on improving nutritional quality.

A range of molecular markers, derived from expressed sequence tags (ESTs) and genomic DNA, has been instrumental in revealing genome relationships and comparing marker-trait associations across various crops [20]. Comparative genomics, especially among major cereal crops like wheat, has illuminated evolutionary relationships and informed crop improvement strategies. Although functional genomics research in wheat has historically lagged behind that of crops such as maize and rice, recent advancements have been significant. Techniques such as RNA interference, TILLING, "expression genetics" have greatly and contributed to mapping expression quantitative trait loci (eQTLs) and understanding the functions of specific genes [21]. These advancements have helped identify candidate genes for targeted traits, aiding both in trait biology comprehension and the development of diagnostic markers for gene cloning and markerassisted selection. To accelerate wheat genome sequencing, the International Genome Research on Wheat (IGROW) initiative was established and has since evolved into the International Genome Sequencing Wheat Consortium

(IWGSC) [22]. This international collaboration is set to enhance genome sequencing efforts and provide a thorough analysis of the wheat genome's structure and function. In light of these developments, Somers identified five key areas wheat improvement research: genetic for mapping, QTL analysis, molecular breeding, association mapping, and software development [23]. These areas are crucial for advancing wheat breeding and genomics, leading to improved crop productivity and sustainability.

2. APPLICATION OF GENOMICS TO MOLECULAR BREEDING OF WHEAT

2.1 Association Mapping in Wheat

The integration of genomics into wheat molecular breeding represents a significant leap forward in modern agriculture. By utilizing advanced genomic tools and techniques, researchers and breeders can significantly improve the efficiency and accuracy of wheat breeding programs. Genomics provides а broad range of methodologies, including quantitative trait loci (QTL) mapping [24], marker-assisted selection (MAS) [25], genomic selection (GS) [26], association mapping, functional genomics, genome sequencing and assembly, and gene editing technologies such as CRISPR-Cas9 [27]. These techniques enable the development of wheat varieties with enhanced yield, quality, resistance to biotic and abiotic stresses, and improved nutritional content [28]. Incorporating genomics into wheat breeding aims to tackle challenges arising from environmental changes. rising food demand, and evolving pest and disease pressures, thereby contributing to global food security and sustainability.

The use of genomics in wheat molecular breeding includes various methodologies. notably association mapping. This highresolution technique for identifying quantitative trait loci (QTL) capitalizes on linkage disequilibrium (LD) and has significant potential for dissecting complex traits [29]. Association mapping offers numerous advantages, as extensively discussed in the literature. In wheat, certain genomic regions are more amenable to LD/association mapping for QTL detection and fine mapping than others. This variability in LD across different chromosome regions highlights the effectiveness of association mapping in uncovering the genetic architecture of wheat traits [30].

2.2 Marker-assisted Selection in Wheat

In recent decades, the discovery of numerous marker-trait associations has advanced the use molecular markers for marker-assisted of selection (MAS) in bread wheat, a technique increasingly adopted worldwide. Major MAS programs are currently active in the USA, Australia, and at CIMMYT in Mexico. In the USA, a wheat MAS consortium, which includes over 20 wheat-breeding programs, was formed to incorporate MAS into public wheat breeding efforts [31]. These programs have enabled the transfer of up to 27 insect and pest resistance genes and 20 alleles linked to improved bread-making and pasta quality into approximately 180 lines tailored to key US production areas [32]. This initiative has led to the release of 45 MAS-derived germplasm lines. Similarly, Australia's program has targeted 20 different traits, including resistance to various abiotic stresses, resulting in the development of enhanced cultivars. MAS has become the preferred approach for selecting crucial agronomic traits, especially where traditional bioassays were costly or inconclusive, as demonstrated by Agriculture Victoria's selection for cereal cyst nematode resistance [33].

MAS has also been incorporated into backcross breeding to introduce QTLs that enhance transpiration efficiency and to perform negative selection against undesirable traits, such as yellow flour color [34]. In Australia, scientists have utilized computer simulations to create cost-effective marker-assisted wheat strategies. combining restricted breeding with doubled backcrossing haploid (DH) technology to reduce breeding costs by up to 40% [35]. This MAS approach has been successfully applied in wheat breeding programs focused on improving quality and rust disease resistance. At CIMMYT, markers linked to 25 different genes controlling traits such as insect pest resistance, protein quality. homoeologous pairing, and other agronomic characteristics are employed in breeding programs develop enhanced to cultivars tabulated in Table 1 [36]. Some of these markers are "perfect markers" derived from the nucleotide sequences of these genes. Future large-scale sequencing of gene-rich regions (GRRs) by the

International Wheat Genome Sequencing Consortium (IWGSC) is anticipated to aid in isolating key genes for generating improved transgenic crops and developing perfect markers for significant agronomic traits to be used in MAS [37].

2.3 CRISPR/Cas9-mediated Knockout and Its Applications in Wheat

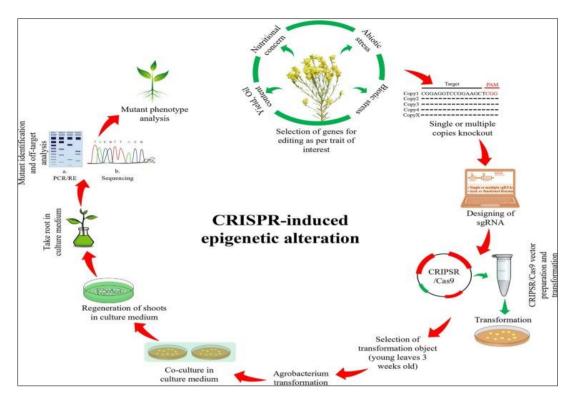
To date, the predominant approach in genomeediting studies has been the use of nonhomologous end joining (NHEJ) to create loss-offunction mutations at specific gene loci in various crops, including wheat. CRISPR/Cas9 has become a widely utilized tool for improving wheat yields and quality (Fig. 1). For example, targeting the lipoxygenase (LOX) gene, which plays multiple roles in plant growth, development, and defense, led to changes in grain size, weight, and improved storability in wheat [38]. Additionally, simultaneous targeting of all three gibberellin-regulated TaGASR7 genes, which affect grain size, resulted in a significant increase in thousand-kernel weight. The deletion of the phosphate 2 gene TaPHO2-A1 enhanced phosphorus uptake and grain yield under low-phosphorus conditions, while knocking out the RING-type E3 ligase gene TaGW2 increased grain size and, consequently, grain yield [39].

Enhancing wheat quality to satisfy varied consumer needs is a central obiective wheat breeding programs. One group of proteins, α-gliadin, gluten of plays а significant role in the end-use properties of wheat in food processing but also contains major immunogenic epitopes that can lead to health issues such as celiac disease [40]. Mutations in genes associated with gliadin, Waxy, and VIT2 have been shown to reduce gliadin content and increase branched starch content. Recently, high-amylose wheat been developed through has targeted mutagenesis of the TaSBElla gene using CRISPR/Cas9, offering potential for creating new wheat varieties with improved nutritional value [41]. Base editing, a precise technique for inducing single-nucleotide point mutations, is also emerging as a valuable method for modifying critical agronomic traits in wheat and other crop plants.

Target gene	Nucleases	Transformation	Improved agronomic traits	Edit knockout	
TaMLO	Cas9	bombardment	powdery mildew resistance		
TaLOX2	Cas9	bombardment	varied grain size, weight and increased storability	knockout	
TaPHO2-A1	Cas9	Agrobacterium	increased phosphorus uptake	knockout	
TaGASR7	Cas9	bombardment	increased yield	knockout	
TaEDR1	Cas9	Agrobacterium	powdery mildew resistance	knockout	
TaGW2	Cas9	bombardment	increased yield	knockout	
TaZIP4	Cas9	Agrobacterium	increased homoeologous CO frequency	knockout	
TaHRC	Cas9	bombardment	Fhb resistance	knockout	
TaMs1	Cas9	bombardment	male sterility	knockout	
TaSBElla	Cas9	bombardment	high amylose	knockout	
TaDA1/TaPDS/ TaNCED1	Cas9	Agrobacterium	_	knockout	
TaCENH3a	Cas9	Agrobacterium	high haploid induction rate	knockout	
TaQsd1,TaARE1, TaNPT1, TaSBEIIa, TaSPDT	Cas9	bombardment	_	knockout and multiplexing	
TaLOX2	nCas9- D10A/ dCas9	Agrobacterium /bombardment	improved wheat quality	base editing	
TaALS, TaACC	nCas9- D10A	bombardment	herbicide resistance	base editing	
Ubi10, TaGW2, TaGASR7, TaDME1 TaLOX2,TaMLO,	nCas9- H840A	protoplast transformation	-	primer editing	
Ubiquitin	Cas9	bombardment	_	HDR replicon	

Table 1. CRISPR/Cas-mediated genome editing in wheat

Source: Bapela et al. [36]



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Fig. 1. Epigenetics for crop improvement using CRISPR [39]

3. QUANTITATIVE TRAIT LOCUS (QTL) MAPPING

Several studies have focused on mapping quantitative trait loci (QTLs) for grain iron (Fe) and zinc (Zn) concentrations in wheat. In 1997, the first QTL associated with iron and zinc concentration, known as Gpc-B1, was identified on chromosome 6BS in a recombinant inbred line (RIL) population derived from a cross between durum wheat and wild emmer (Triticum turgidum) [42]. This QTL was found to increase iron concentration by 18% and zinc concentration by 12%. Numerous QTL studies have been conducted to explore the genomic regions linked to iron and zinc content in wheat grains. Various biparental mapping populations have been employed, primarily using bread wheat cultivars as parents, though other hexaploid species such as Triticum spelta, stabilized synthetic hexaploid wheat (SHW) lines, and tetraploid parents have also been utilized. Additionally, many studies leverage multi-environment trial data to identify stable QTLs for grain Fe and Zn content [43].

In the studies mentioned, QTLs often varied due to differences in populations, environments, and marker sets, which affected the accuracy and positioning of QTL identification. With the advent of new genome sequences, it is now possible to better compare the actual locations of QTLs identified in different studies using these updated reference sequences. This comparison may help in pinpointing QTLs that consistently enhance iron and zinc concentrations. However, practical constraints limit the number of markers that can be used simultaneously for multiple QTLs in large breeding populations, making markerassisted selection challenging. An alternative approach is to use marker-assisted selection to stack small-effect QTLs within the same genomic region, creating a haplotype with multiple beneficial QTLs. A haplotype refers to a set of DNA variations that are inherited together on a single chromosome. By selecting this haplotype as a single unit, the number of markers needed for further selection can be reduced. This method also allows for the integration of QTLs for higher iron and zinc content with those for other desirable traits, such as yield or disease which resistance. are likely already being targeted through marker-assisted selection [44].

Several QTL studies have been conducted to identify genomic regions associated with iron (Fe) and zinc (Zn) concentrations in wheat. For example, a double haploid population derived from a cross between Berkut and Krichauff was analyzed, with grain Zn and Fe concentrations measured using X-ray fluorescence. Composite interval mapping revealed three QTLs: one associated with protein (1A) and two linked to Zn, with one QTL also associated with Fe [45]. These QTLs, identified across multiple seasons, appear to be reliable and useful for markerassisted selection (MAS). In a current study at CIMMYT in Mexico, a line with high grain zinc content from China was crossed with the Mexican bread wheat cultivar Roelfs F2007 to develop a recombinant inbred line (RIL) mapping population for QTL identification. Using the annotated wheat genome sequence, researchers identified 55 potential genes related to Zn and Fe content in the grain [46]. Additionally, four QTLs associated with grain Zn and Fe concentrations were found in a RIL population generated from a cross between the Indian variety 'WH 542' and a Synthetic Hexaploid Wheat (SHW) line [47]. Large-scale implementation of breeding programs to develop nutrient-rich elite wheat lines should be a priority. Developing countries particularly affected by micronutrient are deficiencies, making bio-fortification of wheat with QTLs that enhance grain iron (Fe) and zinc (Zn) content a cost-effective and sustainable solution. To boost Fe and Zn levels, it is essential to screen diverse sources such as unknown landraces. related species, wild relatives, promising synthetic hexaploid wheat (SHW) lines, and alleles from high-nutrient genetic resources. Mapping populations across various environments is critical for understanding genotype-environment interactions and identifying stable QTLs for these traits. Incorporating reported QTLs into elite genetic backgrounds can lead to the development of

nutrient-rich breeding lines, offering a long-term approach to addressing malnutrition [48].

4. GENE CLONING METHODS

After phenotyping a promising mutant, the next critical step is to identify the specific gene responsible for the observed trait. This process relies on genomic sequences as the foundation for gene cloning. Before the advent of reference genomes, many genes associated with important agricultural traits in crops were identified based on their genetic characteristics alone. Positional or map-based cloning is a valuable technique for gene identification. This approach involves creating high-resolution mapping populations that are phenotyped for the desired trait and genotyped using molecular markers. Bv employing mapping populations consisting of several hundred to thousands of plants generated through genetic recombination. researchers can progressively locate the site of a mutant gene using methods like Mutmap-based cloning [46]. The integration of phenotypic and molecular marker data allows for the construction of a genetic map that covers the target locus. Map-based cloning does not require prior knowledge of the gene product to be effective. This technique has successfully cloned several genes, such as Lr10, HM1, and Xa21 from wheat, maize, and rice, respectively [49]. Notably, approximately half of these cloned genes confer disease resistance, as shown in Table 2. However, map-based cloning is laborintensive and time-consuming, and it is not feasible for genes located in centromeric regions [50].

Genes	Function	Identification method	Reference
Lr21	Leaf rust resistance	Map-based cloning	[51-53]
Lr10			
Lr34			
Yr36	Stripe rust resistance	Map-based cloning	[54-55]
Yr10			
Sr33	Stem rust resistance	Map-based cloning	[56-57]
Sr35			
Pm2	Powdery mildew resistance	MutChromSeq	[58]
Fhb1	Fusarium head blight resistance	Map-based cloning	[59]
TaTAR2.1-3A	Boost production and nitrogen usage	Homology-based	[60]
	effectiveness	cloning	
Bo1, Bo4	Boron transporter	Map-based cloning	[61-62]
Gpc-B1	Boosting iron, zinc, and kernel protein by	Map-based cloning	[63]
-	controlling senescence.	-	

Table 2. F	unction o	of t	he g	genes	cloned	in	wheat
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5. FUTURE PERSPECTIVES OF GENOME EDITING

The future trajectory of wheat breeding and genome editing holds immense promise, driven by the ongoing advancements in cutting-edge technologies and innovative strategies outlined in our review. Looking forward, several key developments are expected to shape the field:

1. Advanced Genome Editing Technologies: The refinement and integration of genome editing tools, such as CRISPR/Cas-mediated base prime editina and editina. will offer unprecedented precision and efficiency in modifvina the wheat genome. These technologies will enable the targeted alteration of specific genes, improving traits such as yield, disease resistance, and nutritional quality.

2. Convergence of Genomic Approaches: The synergy of genome-wide association studies (GWAS), marker-assisted selection (MAS), and quantitative trait locus (QTL) mapping will enhance the identification of crucial genes linked to desirable traits. This convergence will accelerate the development of improved wheat varieties by providing comprehensive insights into the genetic basis of key agronomic and nutritional traits.

3. Multiplex Genome Editing: The advent of multiplex genome editing systems will facilitate the simultaneous modification of multiple genes. This capability will enable the pyramiding of beneficial traits, such as improved disease resistance and enhanced nutritional content, into elite wheat cultivars, leading to more robust and high-performing varieties.

4. Genotype-Independent Transformation: Advances in genotype-independent wheat methods will broaden transformation the applicability of genome editing across diverse germplasms. This inclusivity will support the development of wheat varieties suited to a wide of environmental conditions range and agricultural contexts.

5. Synthetic Biology and Gene Stacking: Synthetic biology approaches will offer new opportunities for gene stacking, allowing the accumulation of multiple transgenes to confer complex traits or introduce novel functionalities in wheat. This capability will be particularly valuable for addressing multifaceted challenges in wheat production. 6. Speed Breeding Integration: The integration of speed breeding techniques with rapid advancements in genome editing will expedite breeding cycles. This combination will enable the faster development of new wheat varieties, helping to meet the evolving demands of agriculture and address food security challenges.

7. Collaborative Initiatives and Knowledge Sharing: Collaborative research efforts and knowledge-sharing platforms will play a crucial role in advancing scientific discoveries, promoting the adoption of new technologies, and developing regionally adapted wheat varieties. These initiatives will enhance the collective ability to address local and global agricultural challenges.

In summary, the future of wheat breeding and genome editing is poised to revolutionize agricultural practices. By leveraging these emerging technologies and strategies, we can enhance nutritional quality, improve yields, and promote sustainable practices, ultimately contributing to global food security and agricultural resilience.

6. CONCLUSION

The future of wheat breeding and genome editing is poised for transformative advancements, driven by the integration of cutting-edge technologies and innovative strategies. The adoption of advanced genome editing tools, such CRISPR/Cas-mediated base editing and as prime editing, promises precise and efficient modifications to the wheat genome, ushering in an era of tailored genetic improvements. These technologies will enable the development of elite wheat varieties with enhanced traits, improving resilience to both biotic and abiotic stresses while maximizing productivity and nutritional quality. The use of multiplex genome editing systems and genotype-independent transformation strategies will further accelerate the breeding allowing the simultaneous process by modification of multiple genes and expanding the range of germplasms that can be targeted. This will facilitate the creation of wheat varieties with stacked beneficial traits, addressing critical challenges in modern agriculture. The convergence of diverse genomic approaches, including genome-wide association studies (GWAS), marker-assisted selection (MAS), and quantitative trait locus (QTL) mapping, will provide valuable insights into the genetic basis of complex traits. This understanding will enable

more targeted and effective breeding efforts. leading to the development of varieties that meet specific agricultural and nutritional needs. Synthetic biology methodologies will offer new opportunities for gene stacking and trait diversification, while speed-breeding techniques will shorten breeding cycles, allowing for rapid variety development in response to evolving demands. Collaborative initiatives and knowledge-sharing platforms will play a crucial role in fostering innovation, promoting the adoption of new technologies, and ensuring equitable access to advanced breeding tools. As we face the challenges of a changing climate, growing population, and shifting agricultural landscapes, the synergy between wheat breeding and genome editing offers a beacon of hope for sustainable food production and global food security. By harnessing the power of scientific and technological advancements, we can develop resilient. nutritious. and sustainable environmentally wheat varieties tailored to diverse agroecological contexts, ensuring a brighter future for generations to come.

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Reddy B, Kumar B, Kumar R, Thota H. Analysis of heterotic potential for yield and its contributing traits in Wheat (*Triticum aestivum* L.). International Journal of Environment and Climate Change. 2023; 13(9):388-400.
- Santhoshini A, Dubey N, Avinashe HA, Thonta R, Kumar R. Inheritance studies in segregating population of Bread Wheat (*Triticum aestivum* L.). International Journal of Environment and Climate Change. 2023;13(9):277-87.
- 3. Paudel P, Pandey MK, Subedi M, Paudel P, Kumar R. Genomic approaches for improving drought tolerance in wheat (*Triticum aestivum* L.): A comprehensive

review. Plant Archives. 2024;24(1):1289-1300.

- Ohanenye IC, Emenike CU, Mensi A, Medina-Godoy S, Jin J, Ahmed T, Sun X, Udenigwe CC. Food fortification technologies: Influence on iron, zinc and vitamin A bioavailability and potential implications on micronutrient deficiency in sub-Saharan Africa. Scientific African. 2021;11:e00667.
- Balk J, Connorton JM, Wan Y, Lovegrove A, Moore K L, Uauy C and Shewry PR. 2019. Improving wheat as a source of iron and zinc for global nutrition. Nutrition Bulletin, 44(1), 53-59.
- Gonal B, Doggalli G, Kumar B, Bhushan S, Surekha S, Malathi G, Singh L. Exploring the Future of Plant Breeding: Advancements and Challenges. International Journal of Plant & Soil Science. 2023;35(24):49-55.
- 7. Bhattacharyya P, Pathak H, Pal S, Bhattacharyya P, Pathak H, Pal S. Impact of climate change on agriculture: Evidence and predictions. Climate Smart Agriculture: Concepts, Challenges, and Opportunities. 2020:17-32.
- Namdar R, Karami E, Keshavarz M. Climate change and vulnerability: the case of MENA countries. ISPRS International Journal of Geo-Information. 2021;10(11): 794.
- Hussain B, Akpınar BA, Alaux M, Algharib AM, Sehgal D, Ali Z, Aradottir GI, Batley J, Bellec A, Bentley AR, Cagirici HB. Capturing wheat phenotypes at the genome level. Frontiers in Plant Science. 2022;13:851079.
- Kreitzman M, Toensmeier E, Chan K, Smukler S, Ramankutty N. Perennial staple crops: yields, distribution, and nutrition in the global food system. Frontiers in Sustainable Food Systems. 2020;4:216.
- Nadeem MA, Yeken MZ, Tekin M, Mustafa Z, Hatipoğlu R, Aktaş H, Alsaleh A, Cabi E, Habyarimana E, Zencirci N, Karaköy T. Contribution of Landraces in Wheat Breeding. Wheat Landraces. 2021:215-58.
- 12. Lin G, Zhang K, Peng R, Han Y, Xie J, Li J. Clustered Regularly Interspaced Short Palindromic Repeat (CRISPR)/CRISPR-Associated Endonuclease Cas9–Mediated Homology-Independent Integration for Generating Quality Control Materials for Clinical Molecular Genetic Testing. The

Journal of Molecular Diagnostics. 2018; 20(3):373-80.

- Massel K, Lam Y, Wong AC, Hickey LT, Borrell AK, Godwin ID. Hotter, drier, CRISPR: the latest edit on climate change. Theoretical and Applied Genetics. 2021;134(6):1691-709.
- 14. Raza A, Razzaq A, Mehmood SS, Zou X, Zhang X, Lv Y, Xu J. Impact of climate change on crops adaptation and strategies to tackle its outcome: A review. Plants. 2019;8(2):34.
- Ahmad S, Tang L, Shahzad R, Mawia AM, Rao GS, Jamil S, Wei C, Sheng Z, Shao G, Wei X, Hu P. CRISPR-based crop improvements: a way forward to achieve zero hunger. Journal of Agricultural and Food Chemistry. 2021;69(30):8307-23.
- Kouhen M, García-Caparrós P, Twyman RM, Abdelly C, Mahmoudi H, Schillberg S, Debez A. Improving environmental stress resilience in crops by genome editing: insights from extremophile plants. Critical Reviews in Biotechnology. 2023;43(4):559-74.
- 17. Krishnappa G, Rathan ND, Sehgal D, Ahlawat AK, Singh SK, Singh SK, Shukla RB, Jaiswal JP, Solanki IS, Singh GP, Singh AM. Identification of novel genomic regions for biofortification traits using an SNP marker-enriched linkage map in wheat (*Triticum aestivum* L.). Frontiers in Nutrition. 2021;8:669444.
- Wei X, Qiu J, Yong K, Fan J, Zhang Q, Hua H, Liu J, Wang Q, Olsen KM, Han B, Huang X. A quantitative genomics map of rice provides genetic insights and guides breeding. Nature Genetics. 2021;53(2):243-53.
- Jones N, Ougham H, Thomas H, Pašakinskienė I. Markers and mapping revisited: finding your gene. New phytologist. 2009;183(4):935-66.
- Kage U, Kumar A, Dhokane D, Karre S, Kushalappa AC. Functional molecular markers for crop improvement. Critical reviews in biotechnology. 2016;36(5):917-30.
- 21. Li H, Deng H. Systems genetics, bioinformatics and eQTL mapping. Genetica. 2010;138:915-24.
- Soneji JR, Rao MN, Sudarshana P, Panigrahi J, Kole C. Current status of ongoing genome initiatives. InPrinciples and Practices of Plant Genomics, Volume 3 2016 Apr 19 (pp. 341-389). CRC Press.

- 23. Doerge RW. Mapping and analysis of quantitative trait loci in experimental populations. Nature Reviews Genetics. 2002;3(1):43-52.
- Collard BC, Mackill DJ. Marker-assisted selection: an approach for precision plant breeding in the twenty-first century. Philosophical Transactions of the Royal Society B: Biological Sciences. 2008; 363(1491):557-72.
- Crossa J, Pérez-Rodríguez P, Cuevas J, Montesinos-López O, Jarquín D, De Los Campos G, Burgueño J, González-Camacho JM, Pérez-Elizalde S, Beyene Y, Dreisigacker S. Genomic selection in plant breeding: methods, models, and perspectives. Trends in plant science. 2017; 22(11):961-75.
- 26. Zhu C, Gore M, Buckler ES, Yu J. Status and prospects of association mapping in plants. The plant genome. 2008;1(1).
- 27. Smedley MA, Hayta S, Clarke M, Harwood WA. CRISPR-Cas9 based genome editing in wheat. Current Protocols. 2021;1(3):e65.
- Verma AK, Mandal S, Tiwari A, Monachesi C, Catassi GN, Srivastava A, Gatti S, Lionetti E, Catassi C. Current status and perspectives on the application of CRISPR/Cas9 gene-editing system to develop a low-gluten, non-transgenic wheat variety. Foods. 2021;10(10):2351.
- 29. Xu Y, Li P, Yang Z, Xu C. Genetic mapping of quantitative trait loci in crops. The Crop Journal. 2017;5(2):175-84.
- Lu Y, Zhang S, Shah T, Xie C, Hao Z, Li X, Farkhari M, Ribaut JM, Cao M, Rong T, Xu Y. Joint linkage–linkage disequilibrium mapping is a powerful approach to detecting quantitative trait loci underlying drought tolerance in maize. Proceedings of the National Academy of Sciences. 2010;107(45):19585-90.
- 31. William HM, Trethowan R, Crosby-Galvan EM. Wheat breeding assisted by markers: CIMMYT's experience. Euphytica. 2007; 157:307-19.
- 32. Xu Y, Xie C, Wan J, He Z, Prasanna BM. Marker-assisted selection in cereals: platforms, strategies and examples. Cereal genomics II. 2013:375-411.
- Lema M. Marker assisted selection in comparison to conventional plant breeding. Agric Res Technol. 2018;14:555914.
- Tian J, Chen J, Chen G, Wu P, Zhang H, Zhao Y, Tian J, Chen J, Chen G, Wu P, Zhang H. The Concept and Research Progress of MAS. Genetic Analyses of

Wheat and Molecular Marker-Assisted Breeding, Volume 2: Conditional QTL Analysis and MAS. 2015:179-85.

- 35 Chaikam V, Molenaar W, Melchinger AE, Boddupalli PM. Doubled haploid technology for line development in maize: technical advances and prospects. Theoretical and Applied Genetics. 2019;132:3227-43.
- 36. Bapela T, Shimelis H, Tsilo TJ, Mathew I. Genetic improvement of wheat for drought tolerance: Progress, challenges and opportunities. Plants. 2022;11(10):1331.
- Song L, Wang R, Yang X, Zhang A, Liu D. Molecular markers and their applications in marker-assisted selection (MAS) in bread wheat (*Triticum aestivum* L.). Agriculture. 2023;13(3):642.
- 38. Filip E, Woronko K, Stępień E, Czarniecka N. An overview of factors affecting the functional quality of common wheat (*Triticum aestivum* L.). International journal of molecular sciences. 2023;24(8):7524.
- Soneji JR, Rao MN, Sudarshana P, Panigrahi J, Kole C. Current status of ongoing genome initiatives. InPrinciples and Practices of Plant Genomics, Volume 3 2016 Apr 19 (pp. 341-389). CRC Press.
- Boukid F, Mejri M, Pellegrini N, Sforza S, Prandi B. How looking for celiac-safe wheat can influence its technological properties. Comprehensive Reviews in Food Science and Food Safety. 2017;16(5):797-807.
- 41. Al Rabbi SH, Nadia I, Islam T. CRISPR-Cas Genome Modification for Non-Transgenic Disease-Resistant, High Yielding and High-Nutritional Quality Wheat. InCRISPR and Plant Functional Genomics (pp. 175-188). CRC Press.
- 42. Joppa LR, Du C, Hart GE and Hareland GA. 1997. Mapping gene (s) for grain protein in tetraploid wheat (*Triticum turgidum* L.) using a population of recombinant inbred chromosome lines. Crop Science, 37(5), 1586-1589.
- 43. Gupta PK, Balyan HS, Sharma S and Kumar R. 2021. Biofortification and bioavailability of Zn, Fe and Se in wheat: present status and future prospects. Theoretical and Applied Genetics, 134, 1-35.
- 44. Ali M. W and Borrill P. 2020 Applying genomic resources to accelerate wheat biofortification. Heredity, 125(6), 386-395.
- 45. Tiwari C, Wallwork H, Arun B, Mishra VK, Velu G, Stangoulis J and Joshi AK. 2016.

Molecular mapping of quantitative trait loci for zinc, iron and protein content in the grains of hexaploid wheat. Euphytica, 207, 563-570.

- 46. Keller B, Wicker T, Krattinger SG. Advances in wheat and pathogen genomics: implications for disease control. Annu Rev Phytopathol. 2018;56:67–87.
- Crespo-Herrera LA, Velu G and Singh RP. 2016. Quantitative trait loci mapping reveals pleiotropic effect for grain iron and zinc concentrations in wheat. Annals of Applied Biology, 169(1), 27-35.
- 48. Krishnappa G, Singh AM, Chaudhary S, Ahlawat AK, Singh SK, Shukla RB and Solanki IS. 2017. Molecular mapping of the grain iron and zinc concentration, protein content and thousand kernel weight in wheat (*Triticum* aestivum L.). PLoS One, 12(4), e0174972.
- 49. Johal GS, Briggs SP. Reductase activity encoded by the HM1 disease resistance gene in maize. Science (1979). 1992;258(5084):985–7.
- 50. Feuillet C, Travella S, Stein N, Albar L, Nublat A, Keller B. Map-based isolation of the leaf rust disease resistance gene Lr10 from the hexaploid wheat (*Triticum aestivum* L.) genome. Proceedings of the National Academy of Sciences. 2003;100(25):15253–8.
- 51. Huang L, Brooks SA, Li W, Fellers JP, Trick HN, Gill BS. Map-based cloning of leaf rust resistance gene Lr21 from the large and polyploid genome of bread wheat. Genetics. 2003;164(2):655–64.
- Feuillet C, Travella S, Stein N, Albar L, Nublat A, Keller B. Map-based isolation of the leaf rust disease resistance gene Lr10 from the hexaploid wheat (*Triticum aestivum* L.) genome. Proceedings of the National Academy of Sciences. 2003;100(25):15253–8.
- 53. Krattinger SG, Lagudah ES, Spielmeyer W, Singh RP, Huerta-Espino J, McFadden H, et al. A putative ABC transporter confers durable resistance to multiple fungal pathogens in wheat. Science (1979). 2009;323(5919):1360–3.
- 54. Fu D, Uauy C, Distelfeld A, Blechl A, Epstein L, Chen X, et al. A kinase-START gene confers temperature-dependent resistance to wheat stripe rust. Science (1979). 2009;323(5919):1357–60.
- 55. Liu W, Frick M, Huel R, Nykiforuk CL, Wang X, Gaudet DA, et al. The stripe rust resistance gene Yr10 encodes an

evolutionary-conserved and unique CC– NBS–LRR sequence in wheat. Mol Plant. 2014;7(12):1740–55.

- 56. Periyannan S, Moore J, Ayliffe M, Bansal U, Wang X, Huang L, et al. The gene Sr33, an ortholog of barley Mla genes, encodes resistance to wheat stem rust race Ug99. Science (1979). 2013;341(6147): 786–8.
- 57. Saintenac C, Zhang W, Salcedo A, Rouse MN, Trick HN, Akhunov E, et al. Identification of wheat gene Sr35 that confers resistance to Ug99 stem rust race group. Science (1979). 2013;341(6147): 783–6.
- Sánchez-Martín J, Steuernagel B, Ghosh S, Herren G, Hurni S, Adamski N, et al. Rapid gene isolation in barley and wheat by mutant chromosome sequencing. Genome Biol. 2016;17:1–7.
- 59. Su Z, Bernardo A, Tian B, Chen H, Wang S, Ma H, et al. A deletion mutation in TaHRC confers Fhb1 resistance to

Fusarium head blight in wheat. Nat Genet. 2019;51(7):1099–105.

- 60. Li G, Zhou J, Jia H, Gao Z, Fan M, Luo Y, et al. Mutation of a histidine-rich calciumbinding-protein gene in wheat confers resistance to Fusarium head blight. Nat Genet. 2019;51(7):1106–12.
- Shao A, Ma W, Zhao X, Hu M, He X, Teng W, et al. The auxin biosynthetic TRYPTOPHAN AMINOTRANSFERASE RELATED TaTAR2. 1-3A increases grain yield of wheat. Plant Physiol. 2017;174(4):2274–88.
- Pallotta M, Schnurbusch T, Hayes J, Hay A, Baumann U, Paull J, et al. Molecular basis of adaptation to high soil boron in wheat landraces and elite cultivars. Nature. 2014;514(7520):88–91.
- Uauy C, Distelfeld A, Fahima T, Blechl A, Dubcovsky J. A NAC gene regulating senescence improves grain protein, zinc, and iron content in wheat. Science (1979). 2006;314(5803):1298–301.

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