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Lactic Acid Bacteria Profile of Fermenting Cucumber in 7% Brine Solution

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Authors' contributions

This work was carried out in collaboration between all authors. Authors JGE and JAL designed the study, performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Authors PRA and JGE managed the analyses of the study. Author GEU managed the literature searches. All authors read and approved the final manuscript.

Article Information

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ABSTRACT

Fermentation was used since ancient times as an easy method of vegetables' preservation, which also maintains and/or improves the nutritional and sensory properties of vegetables. The aim of this study was to isolate, characterized and identify the lactic acid bacteria (LAB) in spontaneous fermentation of Cucumber. Cucumber samples were purchased from a vegetable stand in Watt market, Cross River State, Nigeria. They were fermented spontaneously for 7 days in 7% brine solution and samples were taken at 24 h intervals for LAB analysis. Thirty bacterial strains were isolated from different phases of the fermentation of cucumber and were characterized by the API 50 CHL System of identification. Three lactic acid bacteria species were identified and included *Lactobacillus plantarum*, *Leuconostoc mesenteroides*, and *Lactobacillus pentosus*. After 24 h fermentation period. This trend continued until the fourth day where *Lactobacillus plantarum* became the dominant LAB isolate in the fermenting cucumber. As the fermentation proceeded, *Lactobacillus pentosus*, was then involved. The results also revealed a decrease in the occurrence

of *Leuconostoc mesenteroides* as the fermentation period increases. The present research thus shows the presence of LAB at different phases of fermenting cucumber. Therefore, the above mentioned isolates could be used as starter cultures for the commercial fermentation of vegetables.

Keywords: Lactic acid bacteria; cucumber; fermentation; fermented vegetables; starter culture.

1. INTRODUCTION

Mankind has exploited lactic acid bacteria (LAB) for thousands of years for the production of fermented foods because of their ability to produce desirable changes in taste, flavor and texture [1]. Different antimicrobial molecules such as lactic acid, acetic acid, hydrogen peroxide and bacteriocins produced by these bacteria are widely known to inhibit foodborne pathogens and food spoilage micro-organism, thereby extending the shelf-life and enhancing the safety of the food products [1]. The LAB in the fermented foods not only contribute to the taste, aroma and texture but also lower the product's pH, effectively promoting its quality and safety [2]. Lactic acid (LA) fermentation is considered as a simple and useful form of biotechnology to keep and/or enhance the safety, nutritional, sensory and shelf life properties of vegetables and fruits [3].

Cucumber is an important vegetable consumed in Nigeria. Distribution and succession of lactic acid bacteria in cucumber have attracted special attention [4,5]. Lactic acid bacteria are intrinsically present in vegetables, plants materials and dairy products among other foods. Consequently, LAB may be added to minimally processed vegetable product intended for consumption [6]. LAB is natural colonizer of fresh vegetables and has been previously described as good antagonist of several bacteria and fungi in different food products [7]. According to [8], LAB has the potential to inhibit the growth of pathogenic and spoilage bacteria and possibility exist for using them to improve the shelf life of different foods.

The microbiota responsible for the spontaneous fermentation of several raw vegetables and fruits deserves a large interest as tool to improve the microbial safety of fermented foods [9,10]. Biopreservation is mainly due to the synthesis of a wide variety of antagonistic primary and secondary metabolites including organic acids, carbon dioxide, ethanol, hydrogen peroxide and diacetyl, antifungal compounds (e.g., fatty acids, phenyllactic acid), bacteriocins and antibiotics (reutericyclin) [9]. Among these compounds, bacteriocins have attracted the interest due to their potential use as safe and natural food

preservatives. Although nisin is the only purified bacteriocin used thus far in industrially processed foods, many bacteriocins produced by various lactic acid bacteria, may have potential foods. Exploitation applications in of bacteriocinogenic lactic acid bacteria on common spoilage and poisoning of raw vegetables and fruits was largely carried out. Several examples of bio-preservation of fresh cut salads and other vegetables (e.g., apples and lettuce) from common spoilage (yeasts and moulds) and poisoning (Listeria innocua. Listeria monocytogenes and Escherichia coli) microorganisms were reported [9].

Notwithstanding the reliable value of the spontaneous fermentation to stabilize and preserve raw vegetables and fruits (e.g., onions, cucumbers. eggplants, red-beets. capers, lychee, cocoa beans, and persimmon), a number of factors are in favour of the use of selected starters. The risk of fermentation failure. the inadequate inhibition of spoilage and pathogen microorganisms, and the undesirable and not predictable variations of the sensory, nutritional and rheology properties are some of these factors. Contrarily to other fermented foods (e.g., cheeses, sausages and leavened baked goods), the use of starter cultures in vegetable and fruit fermentation is increasing only recently [11,12].

Lactic acid bacteria play important role in food, feed fermentation and preservation either as the natural microflora or as starter culture added under controlled condition. The preservative effect exerted by LAB is mainly due to the production of organic acids (such as lactic acid) which lowered pHs [13,14]. Spontaneous fermentation typically results from the competitive activity of a variety of autochthonous and contaminating microorganisms, which may lead to a high risk for failure. Both from a hygiene and safety perspective, the use of starter cultures is recommended, as it leads to rapid inhibition of spoilage and pathogenic bacteria while yielding processed fruit with consistent sensory and nutritional quality. Starter cultures with desirable properties and high counts of bacteria are of particular importance for the production of high guality fermented vegetable products. However,

it is important to know that, there is few or lack of commercial bacterial cultures suitable for fermentation of vegetable. In view of that, there is need to deepen the search for more potent strain of LAB that can be employed as suitable starter cultures for the fermentation of vegetables. Therefore, this research work isolated, characterized and identified the various LAB involved in the spontaneous fermentation of cucumber.

2. MATERIALS AND METHODS

2.1 Sample Collection and Processing

Fresh cucumber samples were obtained from a vegetable stand in Watt Market, Calabar, Cross River State, Nigeria. They were fermented spontaneously at 37°C for 7 days in 7% brine solution and samples were taken at 24 h interval for microbial analysis.

2.2 Isolation of Lactic Acid Bacteria

After 24 h of fermentation, 0.1ml of 10⁻⁶ dilution of the sample was then spread plated on de Man Rogosa and Sharpe (MRS) agar and incubated at 37°C for 24-48 hrs. Colonies were randomly selected based on their morphological appearance and streak plating was then used to purify the isolates. The same procedure was repeated after every 24 hrs for 7 days. The individual bacterial colonies were stored in MRS agar slant at 4°C for further analysis.

2.3 Identification of Lactic Acid Bacterial Isolates

Isolates were identified using API 50 CHL system (bioMerieux). For primary species- specific identification, bacterial strain were subjected to API 50 CHL assay. Purified strains were cultured on MRS plates, after which grown colonies were cultivated in 5 ml of MRS both at 30°C. The result was read after 24 h and verified after 48 h. Fermentation of carbohydrates in the carbohydrate medium was indicated by a yellow colour, except for esculine (dark brown). Color reactions were scored against a chart provided by the manufacturer. The isolates were also Gram stained to determine their gram's reaction as well as cell morphology and arrangements.

3. RESULTS

Thirty bacterial strains were isolated from different phases of the fermentation of cucumber and were characterized by the API 50 CHL System of identification. The cultural characteristics of the isolates are shown in Table 1. All the thirty isolates reacted positively to gram staining. Morphologically, the cells of 10 isolates were coccus type and arranged either in pairs or tetrads. Their colonies on MRS agar were circular, low convex with entire margin and cream colored. The cells of the remaining 20 were rod shaped and arranged in pairs or chains. Fifteen of the isolates were identified as Lactobacillus plantarum, ten as Leuconostoc mesenteroides, three as Lactobacillus pentosus and two isolates were characterized as Lactobacillus sp as they could not been able to be identified at species level. This result is presented in Table 2. After 24 h fermentation Leuconostoc mesenteroides was found to be the most dominant lactic acid bacteria isolated, followed by Lactobacillus plantarum. This trend continued until the fourth day of fermentation where Lactobacillus plantarum became the dominant lactic acid bacteria in the fermenting cucumber. As the fermentation proceeded, Lactobacillus pentosus was then involved. This agrees with the work of [15]. They showed the different categories of lactic acid bacteria involved in the fermentation of cucumber based on their carbohydrate fermentation capabilities. The heterofermenters initiate the desirable lactic acid fermentation process. This was carried out by *Leuconostoc mesenteroides* as observed from the results. Leuconostoc mesenteroides initiates growth in vegetables more rapidly over a range of temperatures and salt concentrations than any other lactic acid bacteria. It produces carbon dioxide and acids which rapidly lower the pH and inhibit the development of undesirable microorganisms. Once the acidity increases to a specific level, Lactobacillus plantarum takes over the fermentation. It's a homofermenter which only produce lactic acid.



Plate A. Showing different colonies of LAB on MRS agar plate

Egbe et al.; JAMB, 3(1): 1-8, 2017; Article no.JAMB.33457

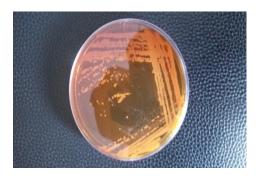


Plate B. Showing discrete colonies of subculture LAB isolate

4. DISCUSSION

Traditionally fermented foods, including fermented vegetables can be a rich source of new LAB strains, with interesting functional properties and with potential applications in food industry and health [16]. Sauerkraut, fermented cucumbers, and kimchi are the most studied lactic acid fermented vegetables mainly due to their commercial importance. The use of starter cultures selected within the autochthonous microbiota of vegetables and fruit is increasing. Compared to commercial LAB, autochthonous strain show better performances during



Plate C. API fermentation kit for LAB isolate C

Egbe et al.; JAMB, 3(1): 1-8, 2017; Article no.JAMB.33457

fermentation and especially ensure higher viability during storage [17]. Fermentation of fruits and vegetables can occur "spontaneously" by the natural lactic bacterial surface microflora, such as Lactobacillus spp, Leuconostoc spp, and Pediococcus spp.; however, the use of starter culture such as L. plantarum, L. rhamnosus, L. gasseri, and L. acidophilus provides consistency and reliability of performance [18]. The present research thus investigated the LAB present at different phases of fermenting cucumber. The result of the study revealed that Lactobacillus plantarum, Leuconostoc mesenteroides and Lactobacillus pentosus were the main LAB involved in the spontaneous fermentation of cucumber. This is in line with the work of [15], were these LAB species were involved in the microbial dynamics of fermenting cucumber. These species particularly Lactobacillus plantarum and Leuconostoc mesenteroides are typical for spontaneous fermentation of vegetables [19]. The result of this study is in accordance with those obtained by [20]. They isolated several lactobacillus strains from fermented vegetables, however, Lactobacillus plantarum was the dominating specie in natural cucumber fermentation. The result also revealed a decrease in the occurrence of Leuconostoc mesenteroides as the fermentation period increases. This could be as a result of acid formation (Lactic acid). This agreed with the work of [21], where they showed that L. plantarum was chiefly responsible for the brine acidity of the fermented cucumbers and that other common types of LAB such as species of Leuconostoc or the gas-producing species of Lactobacillus did not contribute to acid formation in the fermentation. Lactobacillus plantarum isolated in this work is in line with the work of [22], who isolated Lactobacillus plantarum from commercial cucumber fermentation. The ability of some species of LAB particularly Lactobacillus plantarum in acidification of the substrates is significant in food preservation [23]. Consumer

Isolate-code/ fermentation time	Gram's reaction	Pigmentation	Size	Elevation	(Cell-shape) morphology
LD ₁₋ a	+	Creamy	Tiny	Raised	Cocci
LD ₁ _b	+	Creamy	Large	Raised	Cocci
LD ₁₋ c	+	Creamy	Large	Raised	Rod
LD ₂₋ a	+	White	Tiny	Flat	Cocci
LD ₂₋ b	+	Creamy	Tiny	Flat	Rod
LD ₂₋ c	+	Creamy	Large	Flat	Rod
LD ₂₋ d	+	White	Large	Flat	Cocci
LD ₃₋ a	+	White	Large	Raised	Cocci
LD ₃₋ b	+	White	Large	Flat	Cocci
LD ₃₋ c	+	Creamy	Small	Raised	Rod
LD ₃₋ d	+	White	Tiny	Raised	Rod
LD ₃₋ e	+	Creamy	Tiny	Flat	Cocci
LD ₃₋ f	+	Creamy	Small	Pin point	Rod
LD ₄₋ a	+	White	Tiny	Flat	Rod
LD ₄₋ b	+	White	Tiny	Flat	Rod
LD ₄₋ c	+	White	Small	Pin point	Rod
LD ₄₋ d	+	White	Large	Raised	Rod
LD ₄₋ e	+	Creamy	Large	Raised	Rod
LD ₄₋ f	+	Creamy	Small	Flat	Rod
LD₅₋a	+	White	Large	Flat	Cocci
LD ₅₋ b	+	White	Small	Raised	Rod
LD ₅₋ c	+	Creamy	Small	Raised	Rod
LD ₆₋ a	+	White	Large	Pin point	Rod
LD ₆₋ b	+	Creamy	Small	Flat	Rod
LD ₆₋ c	+	White	Small	Flat	Rod
LD ₆₋ d	+	Creamy	Large	Raised	Rod
LD ₆₋ e	+	Creamy	Tiny	Flat	Rod
LD7 ₋ a	+	Creamy	Large	Flat	Rod
LD7_b	+	Creamy	Small	Raised	Rod
LD7.c	+	creamy	Small	Raised	Rod

Table 1.	. Macro	morphological	characteristics	of the isolates
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Sample LAB	Identification	% of reliability of identification
LD ₁₋ a	Leuconostoc mesenteroides	87.5%
LD ₁₋ b	Leuconostoc mesenteroides	98.9%
LD ₁₋ c	Lactobacillus plantarum	99.3%
LD ₂₋ a	Leuconostoc mesenteroides	98.1%
LD ₂₋ b	Lactobacillus plantarum	73.8%
LD ₂₋ c	Leuconostoc mesenteroides	97.2%
LD ₂₋ d	Leuconostoc mesenteroides	90.3%
LD₃₋a	Leuconostoc mesenteroides	99.5%
LD ₃₋ b	Leuconostoc mesenteroides	98.8%
LD ₃₋ c	Leuconostoc mesenteroides	99.0%
LD₃- d	Lactobacillus pentosus	67%
LD₃-e	Lactobacillus plantarum	99.1%
LD ₃₋ f	Leuconostoc mesenteroides	64.5%
LD₄₋a	Lactobacillus plantarum	96.8%
LD ₄₋ b	Lactobacillus pentosus	99.2%
LD ₄₋ c	Lactobacillus plantarum	97.8%
LD ₄₋ d	Lactobacillus petosus	66.7%
LD ₄₋ e	Leuconostoc mesenteroides	98.3%
LD ₄₋ f	<i>Lactobacillus</i> sp	95.4%
LD₅₋a	Lactobacillus plantarum	71.4%
LD ₅₋ b	Lactobacillus plantarum	89.6%
LD ₅₋ c	Lactobacillus plantarum	90.5%
LD ₆₋ a	Lactobacillus plantarum	90.6%
LD ₆₋ b	Lactobacillus plantarum	98.4%
LD ₆₋ c	Lactobacillus plantarum	94.7%
LD ₆₋ d	Lactobacillus plantarum	99.2%
LD ₆₋ e	Lactobacillus plantarum	96.8%
LD7.a	Lactobacillus plantaum	87.5%
LD7 ₋ b	Leuconostoc mesenteroides	96.8%
LD7 ₋ c	Lactobacillus sp	80.7%

Table 2. Phenotypic identification of lactic acid bacteria from fermenting cucumber using API50 CHL gallery kit

interest for diverse fermented foods has increased in recent years because of the positive perception of their beneficial impact on health. Hence, there is an evident need to find novel methods and new food preservation agents from natural origins. Biopreservation refers to extending the shelf-life and enhancing the safety of foods using microorganisms or their metabolites [24]. In this aspect, LAB are very good candidates [25].

5. CONCLUSION

It can be seen that the Lactic acid bacteria involved in the spontaneous fermentation of cucumber are *Lactobacillus plantarum*, *Leuconostoc mesenteroides and Lactobacillus pentosus*. These lactic acid bacteria species can be exploited as starter cultures for the fermentation of other vegetables and fruits.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

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Egbe et al.; JAMB, 3(1): 1-8, 2017; Article no.JAMB.33457

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