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MicroRNAs miR-1 and miR-206 Regulate Monocarboxylate Transporter-4 and Vascular Endothelial Growth Factor Gene Expression in Colorectal Cancer

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Authors' contributions

This work was carried out in collaboration among all authors. Author AK designed the study. Author RAE performed the statistical analysis author CIB wrote the protocol. Author WAR wrote the first draft of the manuscript. Authors AAE, AT and AM managed the analyses of the study. Author YSB managed the literature searches. All authors read and approved the final manuscript.

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ABSTRACT

Background: Colorectal cancer (CRC) is currently the third most common cancer type in males and the second most occurring in females. The role of microRNA (miRNA) in the development of colorectal cancer is not fully elucidated. Therefore, understanding the mechanistic interaction between miRNA and their target oncogenes may hold great importance as a possible target for interventional anticancer therapy.

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Aims: To identify miRNAs that are part of the regulating pathway of Monocarboxylate Transporter-4 (*MCT4*) and Vascular Endothelial Growth Factor (*VEGF*) oncogenes.

Study Design: We used publicly available prediction tools (e.g. TargetScan, MicroCosm, PicTar, and DIANA-microT-CDS) to identify the possible miRNA that target the two oncogenes.

Methodology: We used the GeneMania database to visualize the network and verify gene names and remove ambiguity and duplications. Furthermore, we used miRTarBase database to identify experimentally validated targets which we used to further confirm miRNA-oncogene relationships. Finally, we utilized miR-Mfold web-tool to further visualize the circular structures and the simulated miR-1 and miR-206 targeting arrangements.

Results: We found two putative miRNA (miR-1 and miR-206) that may downregulate MCT4 coded by *SLC16A3* gene and VEGF which is coded by *VEGF gene*. We found relationships between the validated target genes of miR-1 and miR-206 through GeneMania which we extracted from the literature. And we elucidated the proposed structure of these two miRNAs through miR-Mfold webtool.

Conclusion: Our results elucidated a novel regulation pathway in CRC cells and may suggest a potential therapeutic approach for CRC therapy. MiR-1 and miR-206 may help cells go to apoptosis and inhibit the angiogenesis of colorectal cancer cells by down-regulation of MCT4 and VEGF proteins in tumor tissues.

Keywords: MCT4; colorectal cancer; VEGF; microRNA; miR-1; miR-206.

1. INTRODUCTION

Monocarboxvlate transporters (MCTs) are expressed in normal colonic epithelium and facilitate the transport of butyrate, the primary energy source for these cells [1]. However, in colorectal tumor cells, lactate is produced and transported via cell membranes during glycolysis and utilized for energy. The intracellular pH is regulated as the influx and efflux of lactate is controlled by MCTs. MCTs, hence have a vital role in the regulation of pH homeostasis [2]. If this balance is disrupted, the cells normally go through apoptosis. For carcinoma cells to survive by avoiding apoptosis, the control of lactate in glycolysis is considered necessary, and MCTs play an important role in this process [3]. In carcinogenesis, monocarboxylate transporter MCT-4 has a role in the efflux of lactate from tumor cells, which results in escaped apoptosis [4]. Moreover, MCT4 has been reported to be induced by the hypoxic conditions which are usually present in the tumor microenvironment [5].

VEGF is a well-known growth factor and numerous scientific evidences proved the indisputable role of Vascular Endothelial Growth Factor (VEGF) in angiogenesis as well as carcinogenesis [6]. Both MCT4 and VEGF were recently found to be overexpressed in colorectal cancer (CRC) [7].

Micro RNAs, which are usually shortened to miRNAs or miRs, are single-stranded RNAs

capable of posttranscriptional gene regulation via either degrading or suppressing target mRNA [8]. Moreover, in CRC, miRNAs have dual effect possibility; serving either as tumor suppressors or oncogenes depending on their target gene [9,10,11,12]. VEGF has been studied thoroughly and found to be targeted by several miRNAs such as miR-150 [13], miR-195 [14], miR-503[15] miR-195 and miR-378 [16]. However fewer studies focused on miRNAs targeting VEGF in CRC [17,18,19].

To our knowledge MCT4 regulating miRNAs have not been described previously. In the current study, we aim to identify miRNAs that regulate MCT4 and VEGF using the miRNA target prediction web tools because these two genes are over expressed in CRC.

2. METHODS

2.1 Prediction Web Tools used to Identify miRs

We queried four target prediction web tools; (TargetScan7.2) [20], PicTar [21], MicroCosm previously called miRBase [22] and DIANA-microT-CDS [23]). using the gene name *SLC16A3* for MCT4 and *VEGF* to find putative miRs.

2.1.1 miRNA selection based on web tools prediction

We selected two miRNAs as candidates from the results of step 1 for our study. These miRNAs

(miR-1 and miR-206) have never been investigated in CRC but have been reported to be involved in carcinomas.

PubMed search for experimentally validated targets of miR-1 and miR-206.

We did a PubMed database search of the literature looking for experimentally validated targets of miR-1 and miR-206. This search yielded 24 oncogenes that are targeted by miR-1 and miR-206.

2.1.2 Experimentally validated24 oncogenes targets uploaded to GeneMania

We uploaded the 24 oncogenes that we found in step 3 to the GeneMania databases [24] in order to visualize the network and verify gene names and remove ambiguity and duplications.

2.1.3 miRTarBase for miR-1 and miR-206 miRNA-target interactions (MTIs)

We utilized miRTarBase database to download miR-1 and miR-206 MTIs [25].

2.1.4 mfold web-tool to visualize miR-1 and miR-206 structures

We utilized mfold web-tool [26], (http://mfold.rna.albany.edu/), to visualize miR-1

and miR-206 circular structures and show the virtual miR targeting arrangement.

2.1.5 Pathway analysis and identifying overlapping target genes

To discover the 24 genes pathways and overlapping relationships/diseases. We utilized Gene set enrichment analysis (GSEA) [27] web tool. Furthermore, to identify overlapping target genes we used DIANA-microT-CDS web tool [23].

3. RESULTS AND DISCUSSION

Putative miRs from the four web tools algorithm yielded two candidates; miR-1 and miR-206. Consequently, the PubMed search showed 24 validated gene targets for miR-1 and miR-206. The result of the PubMed database search of the literature for experimentally validated targets of miR-1 and miR-206 are summarized in Table 1.

In the next step we uploaded those target genes to GeneMania which resulted in several genegene interactions including; co-expression (57.54%), physical interactions (19.62%), genetic pathway (18.92%), C-localization (3.63%), genetic interactions (0.15%), shared protein domains (0.13%). In addition, GeneMania yielded a genetic network presented in Fig. 1, where genes in black represent our 24 query genes targeted by miR-1 and miR-206.



Fig. 1. GeneMania network for the 24 genes targeted by miR-1 and miR-206

miRNA	Symbol	Gene name	Cancer type	Reference
miRNA-1	MET	met proto-oncogene	Osteocarcinoma/CRC/ rhabdomyosarcoma/	[28,29,30,31,32]
			thyroid carcinoma	
miRNA-1	ETS1	v-ets avian erythroblastosis virus E26 oncogene homolog 1	hepatocellular carcinoma	[33]
miRNA-1	MACC1	metastasis-associated in colon cancer 1	colon cancer	[30]
miRNA-1	PIM1	pim-1 oncogene	lung cancer	[34]
miRNA-1	PIK3CA	phosphoinositide-3-kinase catalytic subunit alpha	non-small cell lung cancer	[35]
miRNA-1	ANXA2	Annexin A2	Glioblastoma	[36]
miRNA-1	SNAI2	snail family zinc finger 2 lung cancer		[37]
miRNA-1	PTMA	prothymosin-α	bladder cancer	[38]
miRNA-1	PNP	purine nucleoside phosphorylase	maxillary sinus squamous cell carcinoma	[39]
miRNA-1	PAX3	paired box 3	Rhabdomyosarcoma	[29]
miRNA-1	CCND2	cyclin D2	Rhabdomyosarcoma	[29,40]
miRNA-1	SRSF9	serine/arginine-rich 9	bladder cancer	[41]
miRNA-1	PNP	purine nucleoside phosphorylase	prostate cancer	[42]
miRNA-1	PTMA	prothymosin-α	nasopharyngeal carcinoma cells	[43]
miRNA-1	PTMA	prothymosin alpha	bladder cancer	[38]
miRNA-1	FN1	fibronectin1	laryngeal squamous carcinoma	[44]
miRNA-1	TAGLN2	transgelin-2	renal cell carcinoma	[45]
miRNA-1	TAGLN2	transgelin-2	head and neck squamous cell carcinoma	[46]
miRNA-1	TAGLN2	transgelin-2	bladder cancer	[47]
miRNA-1	LASP1	LIM and SH3 protein 1	bladder cancer	[45]
miRNA-1	CXCR4	CXC chemokine receptor 4	thyroid carcinoma	[40]
miRNA-1	FOXP1	forkhead box P1	hepatocellular carcinoma	[48,49]
miRNA-1	HDAC4	histone deacetylase 4	hepatocellular carcinoma	[48,50]
miRNA-206	ESR1	estrogen receptor 1	breast cancer	[51]
miRNA-206	MET	met proto-oncogene	papillary thyroid carcinoma	[52]
miRNA-206	MET	met proto-oncogene	Rhabdomyosarcom	[31]
miRNA-206	NOTCH3	notch 3	HeLa cancer cells	[53]
miRNA-206	BAF53A	BAF complex 53 kDa subunit	Rhabdomyosarcoma	[54]
miRNA-206	FOXO3	forkhead box O3	breast cancer	[55]

Table 1. Experimentally validated genetargets of miR-1and miR-206 in various cancers

miRNA	Species (miRNA)	Target gene	Target gene (Entrez ID)	Experiments
hsa-miR-1	Homo sapiens	MYEF2	50804	PAR-CLIP
hsa-miR-1	Homo sapiens	CDK9	1025	Proteomics
hsa-miR-1	Homo sapiens	CEBPA	1050	Luciferase reporter assay
hsa-miR-1	Homo sapiens	MEF2A	4205	Luciferase reporter assay
hsa-miR-1	Homo sapiens	MEF2A	4205	qRT-PCR
hsa-miR-1	Homo sapiens	GATA4	2626	Luciferase reporter assay
hsa-miR-206	Homo sapiens	<i>NOTCH3</i>	4854	Luciferase reporter assay//qRT-PCR//Western blot// Reporter assay
hsa-miR-206	Homo sapiens	<i>NOTCH3</i>	4854	qRT-PCR//Immunohistochemistry//Western blot

Table 2. MiR-1 and miR-206 validated targets that were extracted from miRTarBase web tool

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MiR-206: UGGAAUGUAAGGAAGUGUGUGG

Fig. 2. The structure arrangement of miR-1 and miR-206 as constructed by the Mfold program (http://mfold.rna.albany.edu/) [25]

The miRNA-target interactions (MTIs) that were extracted from miRTarBase database are shown in Table 2.

To further visualize the circular structures and the simulated miR1 and miR2 targeting arrangements, we utilized miR-Mfold web-tool, the resulting structure is shown in Fig. 2.

MCT-4 is frequently deregulated in various cancer cells, it promotes their migration and proliferation and is associated with the level of malignancy and recurrence [56,57,58]. On the other hand, the expression of *VEGF* was reported more frequently in early compared to advanced-stage cancer types. Recent research suggests that *VEGF* is a negative prognostic factor for CRC [59,60]. Gotanda et al. have shown recently that increased *MCT4/VEGF* expression is associated with tumor growth, infiltration, and angiogenesis in their CRC cohort [7].

MiRNAs are small RNAs that have a regulatory effect on their target mRNAs post-

transcriptionally. The effect of these microRNAs is the inhibition of gene expression via either degradation or suppression of target mRNAs (Fig. 3).

Previous research showed that miRNAs could serve as tumor suppressors or oncogenes [9]. Thus miR-1and miR-206 might help in targeting and regulating cancer cell proliferation, migration, and angiogenesis by downregulating the expression of these two genes: *MCT4* and *VEGF*. Fig. 4 illustrates this theory.

In this study, we found two plausible miRNAs which were computationally validated to show that the mRNA of *MCT4* and *VEGF* is a putative target of miR-1 and miR-206, by using publicly available miRNA target prediction webtools.

Pathway analysis by GSEA [27] revealed that most genes overlap were in PI3K/AKT cancer signaling pathway, and diseases of signal transduction by growth factor receptors and second messengers. As shown in Table 3. DIANA-microT-CDS web tool showed overlapping target genes between the two miRNAs with miTG scores of 0.99 (Supplementary Table 1).

These two miRNAs, miR-1 and miR-206, may help in restoring apoptosis pathways by suppressing MCT4 along with inhibiting angiogenesis by targeting *VEGF* [61,62].

Previous study has reported that miR-1 and miR-206 can regulate angiogenesis by targeting and reducing the levels of *Vegf* gene in zebrafish and the knocking down of miR-1and miR-206 increased angiogenesis in the same setting [63]. Recent reports have also shown that both miR-1

and miR-206 were down-regulated in many human cancer types including CRC [64,34,30,65]. Previous reports showed inhibitory roles of miRNAs of MCTs which could reduce tumor cell proliferation [66]. And some studies showed other potential roles of these small RNAs as negative regulators that may lead eventually to growth suppression in some malignancies [67].

In the future, our computational approach needs to be validated by in-vitro and in-vivo expression studies of both miRNAs and target genes in CRC cell-lines i.e. miR-1 and miR-206 and *MCT4/VEGF*.



Fig. 3. schematic organization of microRNA blocking machinery towards target mRNA



Fig. 4. miR-1 and miR-206 act to downregulate the expression of SLC16A3 and VEGF genes, in turn leading to a reduction in tumor growth and angiogenesis

Table 3. GSEA showing PI3K/AKT cancer signaling pathway, and diseases of signal transduction by growth factor receptors and secondmessengers

Gene set name [# Genes (K)]	Description	# Genes in Overlap(k)	p-value	FDR q-value
REACTOME_DISEASES_OF_SIGNAL_TRANSDUCTI CTION_BY_GROWTH_FACTOR_RECEPTORS_AND_S D_SECOND_MESSENGERS [374]	Diseases of signal transduction by growth factor receptors and second messengers	6	5.36 e ⁻⁸	1.2 e ⁻⁴
PID_MYC_ACTIV_PATHWAY [79]	Validated targets of C-MYC transcriptional activation	4	1.18 e ⁻⁷	1.32 e ⁻⁴
REACTOME_PI3K_AKT_SIGNALING_IN_CANCER [102]	PI3K/AKT Signaling in Cancer	4	3.31 e ⁻⁷	2.46 e ⁻⁴
REACTOME_RNA_POLYMERASE_II_TRANSCRIPTI PTION [1375]	RNA Polymerase II Transcription	8	5.4 e ⁻⁷	3.02 e ⁻⁴
REACTOME_INTRACELLULAR_SIGNALING_BY_SE _SECOND_MESSENGERS [304]	Intracellular signaling by second messengers	5	7.09 e ⁻⁷	3.17 e ⁻⁴
REACTOME_DISEASE [1470]	Disease	8	8.95 e ⁻⁷	3.33 e ⁻⁴
PID_ANGIOPOIETIN_RECEPTOR_PATHWAY [49]	Angiopoietin receptor Tie2-mediated signaling	3	2.95 e ⁻⁶	9.42 e ⁻⁴
PID_KIT_PATHWAY [52]	Signaling events mediated by Stem cell factor receptor (c-Kit)	3	3.54 e ⁻⁶	9.88 e ⁻⁴
REACTOME_MET_ACTIVATES_PI3K_AKT_SIGNAL NALING [6]	MET activates PI3K/AKT signaling	2	4.69 e ⁻⁶	1.06 e ⁻³
KEGG_FOCAL_ADHESION [199]	Focal adhesion	4	4.76 e ⁻⁶	1.06 e ⁻³

4. CONCLUSION

MiRNAs are pivotal regulators of gene expression, as they contribute to multiple critical biological processes, including cell proliferation, angiogenesis, and apoptosis. This study could help in deciphering the potential mechanism of acquired regulation of tumor growth and angiogenesis in CRC. In addition, this work sheds light on the involvement of miR-1 and miR-206 in the tumor inhibitory effect by targeting the two oncogenes VEGF/MCT4. Our results elucidated a novel regulatory pathway in CRC cells and could suggest a potential therapeutic approach for CRC. The possibility of metabolic modification of the tumor microenvironment via regulation or manipulation of MCT4 and VEGF may prove to be a promising target for future studies.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Cuff MA, Lambert DW, Shirazi-Beechey SP, Substrate-induced regulation of the human colonic monocarboxylate transporter, MCT1. J Physiol. 2002;539(2): 361-71.
- 2. Halestrap AP. The SLC16 gene family structure, role and regulation in health and disease. Mol Aspects Med. 2013;34(2-3): 337-49.
- 3. Ganapathy V, Thangaraju M, Prasad PD. Nutrient transporters in cancer:

relevance to Warburg hypothesis and beyond. Pharmacol Ther. 2009;121(1):29-40.

- Gerlinger M. et al. Genome-wide RNA interference analysis of renal carcinoma survival regulators identifies MCT4 as a Warburg effect metabolic target. J Pathol. 2012;227(2):146-56.
- Chiche J, Ricci JE, Pouyssegur J. Tumor hypoxia and metabolism -- towards novel anticancer approaches. Ann Endocrinol (Paris). 2013;74(2):111-4.
- Shibuya M. VEGF-VEGFR Signals in Health and Disease. Biomol Ther (Seoul). 2014;22(1):1-9.
- Gotanda Y. et al. Expression of monocarboxylate transporter (MCT)-4 in colorectal cancer and its role: MCT4 contributes to the growth of colorectal cancer with vascular endothelial growth factor. Anticancer Res. 2013;33(7): 2941-7.
- Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. Cell. 2004;116(2):281-97.
- 9. lorio MV, Croce CM, microRNA involvement in human cancer. Carcinogenesis. 2012;33(6):1126-33.
- Toiyama Y. et al. Serum miR-21 as a Diagnostic and Prognostic Biomarker in Colorectal Cancer. J Natl Cancer Inst. 2013;105(12):849-859.
- 11. Chen X. et al. Expression of the tumor suppressor miR-206 is associated with cellular proliferative inhibition and impairs invasion in ERalpha-positive endometrioid adenocarcinoma. Cancer Lett. 2012;314(1): 41-53.
- Zhang J. et al. Putative tumor suppressor miR-145 inhibits colon cancer cell growth by targeting oncogene Friend leukemia virus integration 1 gene. Cancer. 2011; 117(1):86-95.
- Yu ZY. et al. Expression of microRNA-150 targeting vascular endothelial growth factor-A is downregulated under hypoxia during liver regeneration. Mol Med Rep. 2013;8(1):287-93.
- 14. Wang R. et al. MicroRNA-195 suppresses angiogenesis and metastasis of hepatocellular carcinoma by inhibiting the expression of VEGF, VAV2 and CDC42. Hepatology; 2013.
- 15. Zhou B, et al. MicroRNA-503 targets FGF2 and VEGFA and inhibits tumor

angiogenesis and growth. Cancer Lett. 2013;333(2):159-69.

- 16. Deng H. et al. MicroRNA-195 and microRNA-378 mediate tumor growth suppression by epigenetical regulation in gastric cancer. Gene. 2013;518(2):351-9.
- 17. Yang IP. et al. MicroRNA-93 inhibits tumor growth and early relapse of human colorectal cancer by affecting genes involved in the cell cycle. Carcinogenesis. 2012;33(8):1522-30.
- Xu Q. et al. MiR-145 directly targets p70S6K1 in cancer cells to inhibit tumor growth and angiogenesis. Nucleic Acids Res. 2012;40(2):761-74.
- 19. Yamakuchi M, et al. MicroRNA-22 regulates hypoxia signaling in colon cancer cells. PLoS One. 2011;6(5): 20291.
- Lewis BP, CB Burge, Bartel DP. Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. Cell. 2005;120(1):15-20.
- 21. Krek A, et al., Combinatorial microRNA target predictions. Nat Genet. 2005;37(5): 495-500.
- Griffiths-Jones S. et al., miRBase: microRNA sequences, targets and gene nomenclature. Nucleic Acids Res. 2006; 34:140-4.
- 23. Paraskevopoulou MD. et al. DIANA-microT web server v5.0: service integration into miRNA functional analysis workflows. Nucleic Acids Res. 2013;41:169-73.
- 24. Warde-Farley D. et al. The GeneMANIA prediction server: biological network integration for gene prioritization and predicting gene function. Nucleic Acids Res. 2010;38:214-20.
- 25. Hsu SD. et al. miRTarBase update. An information resource for experimentally validated miRNA-target interactions. Nucleic Acids Res. 2014;42:78-85.
- 26. Zuker M. Mfold web server for nucleic acid folding and hybridization prediction. Nucleic Acids Res. 2003;31(13):3406-15.
- 27. Subramanian A, et al. Gene set enrichment analysis: A knowledge-based approach for interpreting genome-wide expression profiles. Proc Natl Acad Sci USA. 2005;102(43):15545-50.

- 28. Novello C. et al. miRNA expression profile in human osteosarcoma: role of miR-1 and miR-133b in proliferation and cell cycle control. Int J Oncol. 2013;42(2):667-75.
- 29. Reid JF. et al. miRNA profiling in colorectal cancer highlights miR-1 involvement in MET-dependent proliferation. Mol Cancer Res. 2012;10(4):504-15.
- Migliore C. et al. MiR-1 downregulation cooperates with MACC1 in promoting MET overexpression in human colon cancer. Clin Cancer Res. 2012;18(3):737-47.
- Yan D. et al. MicroRNA-1/206 targets c-Met and inhibits rhabdomyosarcoma development. J Biol Chem. 2009;284(43): 29596-604.
- 32. Yip L. et al. MicroRNA signature distinguishes the degree of aggressiveness of papillary thyroid carcinoma. Ann Surg Oncol. 2011;18(7):2035-41.
- Wei W. et al. MicroRNA-1 and microRNA-499 downregulate the expression of the ets1 proto-oncogene in HepG2 cells. Oncol Rep. 2012;28(2):701-6.
- Nasser MW, et al., Down-regulation of micro-RNA-1 (miR-1) in lung cancer. Suppression of tumorigenic property of lung cancer cells and their sensitization to doxorubicin-induced apoptosis by miR-1. J Biol Chem. 2008;283(48):33394-405.
- 35. Yu QQ. et al. MiR-1 targets PIK3CA and inhibits tumorigenic properties of A549 cells. Biomed Pharmacother. 2014;68(2):155-61.
- Bronisz A. et al. Extracellular vesicles modulate the glioblastoma microenvironment via a tumor suppression signaling network directed by miR-1. Cancer Res. 2014;74(3):738-50.
- 37. Tominaga E. et al. MicroRNA-1 targets Slug and endows lung cancer A549 cells with epithelial and anti-tumorigenic properties. Exp Cell Res. 2013;319(3):77-88.
- Yamasaki T. et al. Novel molecular targets regulated by tumor suppressors microRNA-1 and microRNA-133a in bladder cancer. Int J Oncol. 2012;40(6):1821-30.
- Nohata N. et al. Identification of novel molecular targets regulated by tumor suppressive miR-1/miR-133a in maxillary

sinus squamous cell carcinoma. Int J Oncol. 2011;39(5):1099-107.

- 40. Leone V. et al. MiR-1 is a tumor suppressor in thyroid carcinogenesis targeting CCND2, CXCR4, and SDF-1alpha. J Clin Endocrinol Metab. 2011;96(9):1388-98.
- Yoshino H. et al. Tumor suppressive microRNA-1 mediated novel apoptosis pathways through direct inhibition of splicing factor serine/arginine-rich 9 (SRSF9/SRp30c) in bladder cancer. Biochem Biophys Res Commun. 2012;417(1):588-93.
- 42. Kojima S. et al. Tumour suppressors miR-1 and miR-133a target the oncogenic function of purine nucleoside phosphorylase (PNP) in prostate cancer. Br J Cancer. 2012;106(2):405-13.
- 43. Wu CD. et al. MicroRNA-1 induces apoptosis by targeting prothymosin alpha in nasopharyngeal carcinoma cells. J Biomed Sci. 2011;18:80.
- 44. Wang F. et al. miRNA-1 targets fibronectin1 and suppresses the migration and invasion of the HEp2 laryngeal squamous carcinoma cell line. FEBS Lett. 2011;585(20):3263-9.
- Chiyomaru T. et al., Functional role of LASP1 in cell viability and its regulation by microRNAs in bladder cancer. Urol Oncol. 2012;30(4):434-43.
- 46. Nohata N. et al. miR-1 as a tumor suppressive microRNA targeting TAGLN2 in head and neck squamous cell carcinoma. Oncotarget. 2011;2(1-2):29-42.
- Yoshino H. et al. The tumour-suppressive function of miR-1 and miR-133a targeting TAGLN2 in bladder cancer. Br J Cancer, 2011;104(5):808-18.
- 48. Datta J. et al. Methylation mediated silencing of MicroRNA-1 gene and its role in hepatocellular carcinogenesis. Cancer Res. 2008;68(13):5049-58.
- 49. Katoh M. et al., Cancer genetics and genomics of human FOX family genes. Cancer Lett. 2013;328(2):198-206.
- 50. Singh A. et al. Transcription factor NRF2 regulates miR-1 and miR-206 to drive tumorigenesis. J Clin Invest. 2013;123(7):2921-34.
- 51. Adams BD, Cowee DM, White BA. The role of miR-206 in the epidermal growth factor (EGF) induced repression of estrogen receptor-alpha (ERalpha)

signaling and a luminal phenotype in MCF-7 breast cancer cells. Mol Endocrinol. 2009;23(8):1215-30.

- 52. Liu X. et al. Expression profiles of microRNAs and their target genes in papillary thyroid carcinoma. Oncol Rep. 2013;29(4):1415-20.
- 53. Song G, Zhang Y, Wang L. MicroRNA-206 targets notch3, activates apoptosis, and inhibits tumor cell migration and focus formation. J Biol Chem. 2009;284(46):31921-7.
- 54. Taulli R. et al. Failure to downregulate the BAF53a subunit of the SWI/SNF chromatin remodeling complex contributes to the differentiation block in rhabdomyosarcoma. Oncogene; 2013.
- 55. Di Leva G. et al. MicroRNA cluster 221-222 and estrogen receptor alpha interactions in breast cancer. J Natl Cancer Inst. 2010;102(10):706-21.
- Vegran F. et al. Lactate influx through the endothelial cell monocarboxylate transporter MCT1 supports an NFkappaB/IL-8 pathway that drives tumor angiogenesis. Cancer Res. 2011;71(7):2550-60.
- 57. Gallagher SM. et al. Monocarboxylate transporter 4 regulates maturation and trafficking of CD147 to the plasma membrane in the metastatic breast cancer cell line MDA-MB-231. Cancer Res. 2007;67(9):4182-9.
- Pertega-Gomes N. et al. Monocarboxylate transporter 4 (MCT4) and CD147 overexpression is associated with poor prognosis in prostate cancer. BMC Cancer. 2011;11:312.
- Harada Y, Ogata Y, Shirouzu K. Expression of vascular endothelial growth factor and its receptor KDR (kinase domain-containing receptor)/Flk-1 (fetal liver kinase-1) as prognostic factors in human colorectal cancer. Int J Clin Oncol. 2001;6(5):221-8.
- 60. Bendardaf R. et al. VEGF-1 expression in colorectal cancer is associated with disease localization, stage, and long-term disease-specific survival. Anticancer Res. 2008;28(6):3865-70.
- 61. Xu W. et al. MiR-1 suppresses tumor cell proliferation in colorectal cancer by inhibition of Smad3-mediated tumor glycolysis. Cell Death Dis. 2017;8(5): 2761.

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- 62. Xu Z. et al. CCL19 suppresses angiogenesis through promoting miR-206 and inhibiting Met/ERK/Elk-1/HIF-1alpha/VEGF-A pathway in colorectal cancer. Cell Death Dis. 2018;9(10):974.
- 63. Stahlhut C. et al. miR-1 and miR-206 regulate angiogenesis by modulating VegfA expression in zebrafish. Development. 2012;139(23): 4356-64.
- 64. Suzuki T. et al. miR-122a-regulated expression of a suicide gene prevents hepatotoxicity without altering antitumor effects in suicide gene therapy. Mol Ther. 2008;16(10):1719-26.
- 65. Parasramka MA. et al. A role for lowabundance miRNAs in colon cancer: the miR-206/Kruppel-like factor 4 (KLF4) axis. Clin Epigenetics. 2012;4(1):16.
- 66. Pullen TJ. et al. miR-29a and miR-29b contribute to pancreatic beta-cell-specific silencing of monocarboxylate transporter 1 (Mct1). Mol Cell Biol. 2011;31(15):3182-94.
- 67. Li KK. et al. miR-124 is frequently down-regulated in medulloblastoma and is a negative regulator of SLC16A1. Hum Pathol. 2009;40(9):1234-43.

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