



Computational *DIO2* rSNP Analysis, Transcriptional Factor Binding Sites and Disease

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Author's contribution

The sole author designed, analyzed and interpreted and prepared the manuscript.

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Short Research Article

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ABSTRACT

Purpose: The *DIO2* gene transcribes the deiodinase type 2 enzyme that changes the thyroid prohormone, thyroxine (T₄), to the biologically active triiodothyronine (T₃) hormone. T₃ plays a vital part in the regulation of energy balance and glucose metabolism. *DIO2* single-nucleotide polymorphisms (SNPs) were computationally examined with respect to changes in putative transcriptional factor binding sites (TFBS) and these changes were discussed in relation to human disease.

Methods: The JASPAR CORE and ConSite databases were instrumental in identifying the TFBS. The Vector NTI Advance 11.5 computer program was employed in locating all the TFBS in the *DIO2* gene from 2.4 kb upstream of the transcriptional start site to 508 bp past the 3'UTR. The JASPAR CORE database was also involved in computing each nucleotide occurrence (%) within the TFBS.

Results: Regulatory SNPs (rSNPs) in the promoter region novel SNP (-2035bp), 5'UTR (rs12885300), intron one (rs225010, 225011 and rs225012), exon two [rs225014 (Thr92Ala)] and 3' UTR (rs6574549, rs225015 and rs225017) of the *DIO2* gene are in linkage disequilibrium. These rSNP alleles were found to alter the DNA landscape for potential transcriptional factors (TFs) to attach resulting in changes in TFBS.

Conclusion: The alleles of each rSNP were found to generate unique TFBS resulting in potential

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changes in TF *DIO2* regulation. These regulatory changes were discussed with respect to changes in human health resulting in disease or sickness.

Keywords: *DIO2*; rSNPs; TFBS; disease.

1. INTRODUCTION

The type 2 deiodinase gene (*DIO2*) encodes a deiodinase that converts the thyroid prohormone, thyroxine (T₄), to the biologically active triiodothyronine (T₃) hormone where T₃ is involved in the vital role of regulating energy balance and glucose metabolism [1-4]. *DIO2* is found in the thyroid gland, cardiac and skeletal muscle, brown adipose tissue, placenta, pituitary, central nervous system (CNS) and at low levels in kidney and pancreases [5-7]. The *DIO2* gene maps to human chromosome 14q24.3 and is about 15 kb in size. The coding region consists of two exons separated by a gap of approximately 7.4 kb [8]. Several single nucleotide polymorphisms (SNPs) have been found in the gene which have been studied is association with mental retardation (MR) [9], osteoarthritis [10], early-onset type 2 diabetes mellitus (T2DM) [11] and insulin resistance (IR) [12,13]. Three of the common SNPs in the gene (rs225014, rs225012 and rs225010) have been found to be in strong linkage disequilibrium (LD) with each other while the rs225012 and rs225010 SNPs have been shown to have a positive association with MR [9]. The haplotypes of two SNPs (rs225014 and rs12885300) have been shown to have a significant association with symptomatic osteoarthritis in Dutch women [10]. Three SNPs (rs225011, rs225014 and rs225015) which are in LD were found to be modestly associated with early-onset of T2DM in Pima Indians while these SNPs and rs6574549 were found to be nominally associated with hepatic glucose output [11]. Two of the SNPs (rs225014 and rs225017) which are in partial LD have been found to be associated with IR in Caucasian T2DM patients [13]. The rs6574549 SNP was also found to be associated with fasting insulin, insulin action and energy expenditure [11]. These studies suggest that some *DIO2* SNPs may be affecting the regulatory network for the gene expression in humans. When LD is found between SNPs in a gene's regulatory region, it can result from strong associations of certain haplotypes with sickness or disease [14-16]. Consequently, a computational examination was made between *DIO2* SNPs in LD and the transcription factor binding site (TFBS) changes resulting from the SNPs. In this report LD is

considered to be the non-random association of SNP alleles within the gene.

Nucleotide changes that influence gene expression by altering gene regulatory sequences such as in promoters, enhancers, and silencers are known as regulatory SNPs (rSNPs) [17-20]. A rSNP within a transcriptional factor's (TF) ability to bind the motif [21-24] in which case the TF would not effectively regulate the gene [25-29]. This concept is examined for the above *DIO2* rSNPs and their allelic association with TFBS, where computation analyses [30-33] was used to identify TFBS alterations created by the *DIO2* rSNPs. In this study, the rSNP associations with nucleotide substitutions in putative TFBS are examined with their possible relationship to disease in humans.

2. METHODS

The JASPAR CORE database [34,35] and ConSite [36] were used to identify the potential *DIO2* TFBS in this study. JASPAR is a database of transcription factor DNA-binding preferences used for scanning genomic sequences where ConSite is a web-based tool for finding cis-regulatory elements in genomic sequences. The TFBS and rSNP location within the binding sites have previously been discussed [14,16,37]. The Vector NTI Advance 11.5 computer program (Invitrogen, Life Technologies) was used to locate the TFBS in the *DIO2* gene (NCBI Ref Seq NM_013989) from 2.4 kb upstream of the transcriptional start site to 508 bp past the 3'UTR which represents a total of 16.9 kb. The JASPAR CORE database was also used to calculate each nucleotide occurrence (%) within the TFBS, where upper case lettering indicate that the nucleotide occurs 90% or greater and lower case less than 90%. The occurrence of each SNP allele in the TFBS is also computed from the database (Table & Supplement).

3. RESULTS

3.1 *DIO2* rSNPs and TFBS

The *DIO2* gene transcribes the deiodinase type 2 enzyme that changes the thyroid prohormone, thyroxine (T₄), to the biologically active

triiodothyronine (T3) hormone. The thyroid hormones play an important role in energy homeostasis and glucose metabolism. Due to the importance of this gene in energy homeostasis, *DIO2* SNPs associated with disease were computationally evaluated with regard to TFBS. The novel -2035bp SNP is located 5' upstream from the TSS, the rs12885300 SNP is located in the 5'UTR and the rs225010, rs225011 and rs225014 SNPs are found in intron one. The rs224014 (Thr92Ala) SNP is located in exon two while the rs674549, rs225015 and rs225017 SNPs are located in the 3' UTR. The novel -2035bp, rs225011, rs225014, rs674549 and rs225015 SNPs are all in LD with each other [11]. The rs225010 and rs225012 SNPs are also in LD but not with the other SNPs [9]. The novel -2035bp and rs674549 SNPs have very rare alleles with a frequency of 0.004 and 0.007, respectively. Since the minor allele frequencies (MAF) of the other SNPs are rather large ranging from 0.229 to 0.421, the minor alleles that alter BS which give rise to different TFs would be expected to have an impact on *DIO2* regulation (Table).

The *DIO2* SNPs (rs225010 and rs225012) which are in LD have been found to be significantly associated with MR in Chinese [9]. The common rs225010 SNP *DIO2*-C allele creates six unique TFBS for the ELF5, ELK1, GATA2, GATA4, JUN:FOS and SREBF1 TFs, which are involved with the ETS transcriptional factor family, the ras-raf-MAPK signaling cascade, the proliferation of hematopoietic and endocrine cell lineages, myocardial differentiation and function, myocardogenic gene expression, and lipid homeostasis, respectively (Table, supplement). The minor *TIO2*-T allele creates four unique TFBS for the HOXA9, JUND, NFATC2 and ZNF354C TFs which are involved with development, transcription enhancement, inducible expression of cytokine genes in T-cells and transcription repression, respectively (Table, supplement). There are also five conserved TBFS for the GATA1, KLF5, MAFB, NFE2L1:MAFG and PAX TFs which are involved with erythroid development, transcription activation, up-regulation of cytoprotective genes, cell differentiation of erythrocytes and kidney cell differentiation, respectively (Table, supplement). The common rs225012 SNP *DIO2*-G allele creates seven unique TFBS for the E2F6, EGR1, ELF1, ERG, SP1, SPI1 and ZNF263 TFs which are involved with control of cell cycle and action of tumor suppressor proteins, mitogenesis and differentiation,

transcription enhancement and repression, regulator of embryonic development, activation or repression of transcription, myeloid and B-lymphoid cell development, and transcription repression, respectively (Table, supplement). The minor rs225012 SNP *DIO2*-A allele creates eight unique TFBS for the EHF, ELF5, EN1, HLTF, HOXA5, NKX3-2, PDX1 and PRRX2 TFs which are involved with epithelial-specific expression, controlling development, altering chromatin structure, cell development, negative regulator of chondrocyte maturation, glucose-dependent regulation of insulin gene transcription, and proliferating fetal fibroblasts, respectively (Table, supplement). There are also two conserved TBFS for the FOXC1 and SPIB TFs which are involved with regulation of cell viability and resistance to oxidative stress as well as transcriptional activation, respectively (Table, supplement).

Two *DIO2* SNPs (rs225014 and rs12885300) have been shown to have a significant association with symptomatic osteoarthritis in Dutch women [10] while a third SNP (rs225017) has been found to be significantly associated with IR [13]. The rs225014 SNP results in a non-synonymous amino acid substitution (Thr92Ala) in exon 2 and has also been associated with IR in obese Caucasian women [12]. The common rs225014 SNP *DIO2*-T allele creates five unique TFBS for the FOXC1, HOXA5, SPI1, STAT5A:STAT5B and THAP1 TFs which are involved with cell viability and resistance to oxidative stress, cell development and myeloid and B-lymphoid cell development, signal transduction and activation of transcription, and G1/S cell-cycle progression respectively (Table, supplement). The minor rs225014 SNP *DIO2*-C allele creates six unique TFBS for the EBF1, NKX3-2, SP2, SPIB, TFAP2C and ZNF354C TFs which are involved with transcription activation, negative regulation of the chondrocyte maturation, activation of mRNA synthesis and transcription repression, lymphoid-specific enhancement, respectively (Table, supplement). There are also four conserved TFBS for the EGR1, ELF1, HLTF and RXR::RAR_DR5 TFs which are involved with mitogenesis and differentiation, enhancement and repression, altering chromatin structure, regulation of development, respectively (Table, supplement). The common rs12885300 *DIO2*-C allele creates no unique TFBS for TFs. The minor rs12885300 *DIO2*-T alleles creates six unique TFBS for the ARID3A, BATF:JUN, IRF1, JUN:FOS, PAX2 and SOX6 TFs which are involved with B-cell

differentiation, negative regulation of AP-1/ATF transcription events, regulation of cellular responses, signal transduction, cell proliferation and differentiation, kidney cell differentiation and maintenance of cardiac and skeletal muscle cells, respectively (Table, supplement). There are also four conserved TFBS between the rs12885300 SNP alleles for the FOXP1, PRDM1, SOX3 and TCF7L2 TFs which are involved with lung epithelium, repression of beta-interferon gene expression, neuronal development and blood glucose homeostasis, respectively (Table, supplement). The common rs225017 *DIO2*-T allele creates six unique potential TFBS for the HOXA5, JUND(var.2), NFE2L1::MAFG, PDX1, SOX3 and STAT3 TFs which are involved with development regulation, enhancer binding, erythrocyte development, insulin activation, neuronal development and signal transduction (Table, supplement). The minor rs225017 *DIO2*-A allele creates five unique potential TFBS for the CEBPA & B, NKX2-5, PRRX2 and SRY TFs which are involved enhancer binding, inflammation, hemopoiesis, chondrocyte maturation, fetal fibroblasts and male development (Table, supplement). There are also three conserved TFBS between the rs225017 SNP alleles for the BATF::JUN, GATA4 and HAND1:TCFE2L TFs which are involved with negative regulation of AP-1/ATF transcription events, myocardial differentiation and function as well as B lymphopoiesis (Table, supplement).

Three *DIO2* SNPs (rs225011, rs225014 and rs225015) have been modestly associated with early-onset T2DM in Pima Indians [11] while the rs225014 and rs225017 SNPs have been found to be associated with IR in Caucasian morbidly obese subjects and T2DM patients [12,13]. The common rs225011 SNP *DIO2*-C allele creates two unique TFBS for the CRX and RXRA TFs which are involved with photoreceptor cells and retinoic acid-mediated gene activation, respectively (Table, supplement). The minor rs225011 SNP *DIO2*-T allele creates three unique TFBS for the FOXL1, MEF2A and PDX1 TFs which are involved with metabolism, cell proliferation and gene expression, skeletal and cardiac muscle development, glucose-dependent regulation of insulin gene transcription, respectively (Table, supplement). There are also eight conserved TFBS between the rs125011 SNPs alleles for the ESRRA, ESRRB, GATA4, NKX2-5, NR5A2, PRRX2, RORA_1 and RORA_2 TFs which are involved with site-specific transcription regulation, myocardial

differentiation and function, negative regulation of chondrocyte maturation, regulation of cholesterol expression in liver, proliferating fetal fibroblasts, and nuclear hormone receptors, respectively (Table, supplement).

The common rs225015 SNP *DIO2*-G allele creates five unique TFBS for the EBF1, ESRRA, PPARG:RXRA, RFX5 and THAP1 TFs which are involved with transcription activation, site-specific transcription regulation, regulation of adipocyte differentiation and glucose homeostasis, and regulation of endothelial cell proliferation and G1/S cell-cycle progression, respectively (Fig. 1, Table and supplement). The minor rs225015 SNP *DIO2*-A allele creates nine unique TFBS for the ELF1, ELK1, ERG, ETS1, FLI1, RUNX1, SOX9, SPI1 and TCF7L2 TFs which are involved with lymphoid cells, ras-raf-MAPK signaling cascade, regulation of embryonic development, TTRAP, UBE2I and Death associated proteins, transcription activation, normal hematopoiesis, skeletal development, myeloid and B-lymphoid cell development and blood glucose homeostasis, respectively (Table, supplement). There are also ten conserved TFBS between the SNPs alleles for the BRCA1, ELF5, HLTF, NFATC2, NFKB1, SOX2, SOX3, SOX6, SOX10 and SPIB TFs which are involved with genomic stability, epithelium cells, altering chromatin structure, cytokine genes in T-cells, signal transduction, regulation of embryonic development, neuronal development, central nervous system, and lymphoid-specific enhancement, respectively (Table, supplement).

Four *DIO2* SNPs (rs225011, rs225014, rs225015 and rs6574549) were nominally associated with hepatic glucose output while the rs6574549 SNP was also associated with fasting insulin, insulin action and energy expenditure in Pima Indians [11]. The common rs6574549 SNP *DIO2*-T allele creates five unique TFBS for the ARID3A, HNF1B, HOXA5, LHX3 and NKX3-2 TFs which are involved with cell cycle progression, embryonic pancreas development, specific positional identities of cells, pituitary development and chondrocyte maturation, respectively (Table, supplement). The minor rs6574549 SNP *DIO2*-G allele creates four unique TFBS for the FOXA1, FOXA2, NFIL3 and POU2F2 TFs which are involved with embryonic development, expression of interleukin-3, POU domain family, respectively (Table, supplement). There are also seven conserved TFBS between the SNP alleles for the FOXC1, FOXD3, FOXI1, FOXL1, HLTF, NKX3-1 and NKX2-5 TFs which

are involved with cell viability and resistance to oxidative stress, activation and repression, normal hearing, sense of balance and kidney function, ontogenesis, altering chromatin structure, epithelial cell growth, and chondrocyte maturation, respectively (Table, supplement). These four *DIO2* SNPS and a novel SNP in the 5'UTR flanking region were found to be in LD in the Pima Indian study [11]. The common novel SNP *DIO2*-C allele creates four unique TFBS for the BRCA1, NFYA, RUNX1 and RUNX2 TFs which are involved with genomic stability, stimulation of transcription of many genes, development of normal hematopoiesis and maturation of osteoblasts, respectively (Table, supplement). The rare novel SNP *DIO2*-T allele creates nine unique TFBS for the ARID3A, CDX2, GFI1, HOXA9, NKX2-5, NOBOX, PBX1,

SOX17 and STAT3 TFs which are involved with cell cycle progression, cell growth and differentiation, hematopoiesis and oncogenesis, the developmental regulatory system, negative regulation of chondrocyte maturation, oogenesis, glucose-dependent regulation of insulin gene transcription, transcription repression, and cellular responses to interleukins, respectively (Fig. 2, Table, supplement). There are also seven conserved TFBS between the SNP alleles for the FOXD3, FOXI1, FOXQ1, MEIS1, NFYB, SRY and THAP1 TFs which are involved with transcriptional activation and repression, kidney function, follicle differentiation, normal development, binding CCAAT motifs in the promoter, male development and regulation of endothelial cell proliferation, respectively (Table, supplement).

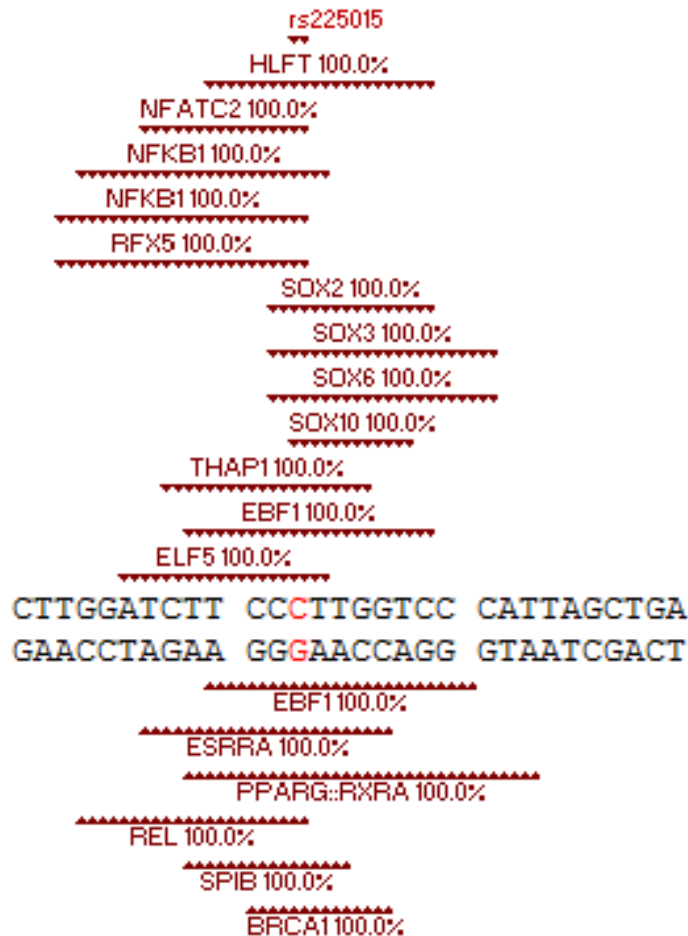


Fig. 1. Double stranded DNA from the *DIO2* 3'UTR showing the potential TFBS for eighteen different TFs which can bind their respective DNA sequence either above (+) or below (-) the duplex (cf. Table). The rs225015 common *DIO2*-G allele is found in each of these TFBS. As shown, this rSNP is located in the 3'UTR of the *DIO2* gene. Also included with the potential TFBS is their % sequence homology to the duplex

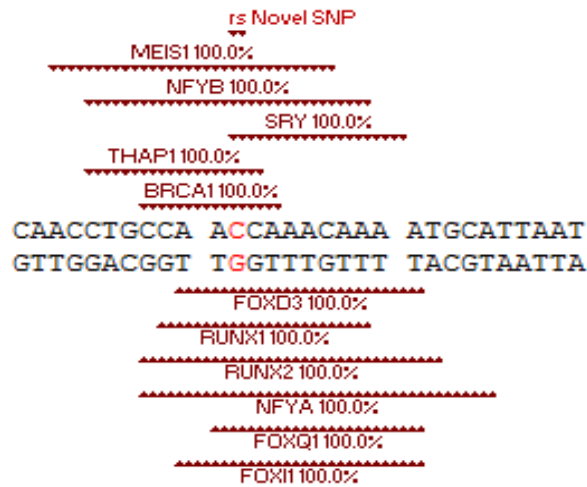


Fig. 2. Double stranded DNA from the DIO2 5'UTR showing the potential TFBS for eleven different TFs which can bind their respective DNA sequence either above (+) or below (-) the duplex (cf. Table). The novel -2035bp rSNP common DIO2-C allele is found in each of these TFBS. As shown, this rSNP is located in the 5'UTR of the DIO2 gene. Also included with the potential TFBS is their % sequence homology to the duplex

4. DISCUSSION

The genome-wide association studies (GWAS) has over the past decade provided us with nearly 6,500 disease or trait-predisposing SNPs. Only seven percent of these SNPs are located in protein-coding regions of the genome [38,39] while the remaining 93% are located within non-coding regions [40,41] such as gene regulatory or intergenic areas of the genome. Much attention has been drawn to SNPs that occur in the putative regulatory of a gene where a single nucleotide change in the DNA sequence of a potential TF motif may affect the process of gene regulation [17,19,42]. A nucleotide change in a TFBS can have multiple consequences. Since a TF can usually recognize a number of different binding motifs in a gene, the SNP may not change the TFBS interaction with the TF and consequently not alter the process of gene expression. In other cases the nucleotide change may increase or decrease the TF's ability to bind DNA which would result in allele-specific gene expression. In some cases a nucleotide change may eliminate the natural binding motif or generate a new BS as a result the gene is no longer regulated by the original TF [14,16]. Therefore, functional rSNPs in TFBS may result in differences in gene expression, phenotypes and susceptibility to environmental exposure [42]. Examples of rSNPs associated with disease susceptibility are numerous and several reviews have been published [42-45].

The rs225012 rSNP *DIO2*-G allele [G (- strand) or C (+ strand)] located in the E2F6 and ELF1 TFBSs have a 100% occurrence in humans while the EGR1 and SPI1 TFBSs have a 94% and 92% occurrence, respectively (Table). Since these binding sites (BS) occurs only once in the gene, this rSNP would probably have a major impact on these TFs regulating the gene. The ERG and SP1 TFBSs also have a 100% occurrence in humans but these BS occur more than once in the gene and should be the rSNP would not have much of an impact gene regulation (Table). The alternate rs225012 rSNP *DIO2*-A allele [A (- strand) or T (+ strand)] located in the PRRX2 TFBS has a 100% occurrence in humans but occurs 55 times in the gene therefore the rSNP would not be expected to have an impact on the TFs regulating the gene (Table). On the other hand, the rs225012 rSNP *DIO2*-A allele located in HOXA5 and NKX3-2 TFBSs also have a 100% occurrence in humans and occur only once in the gene and therefore, should have a major impact on gene regulation since these BS only occur with the minor allele (Table). The E2F6 TFBS provided by the rs225012 rSNP common G allele and not present with the minor A allele is a BS for a TF which is involved with the control of the cell cycle and the action of tumor suppressor proteins. Consequently individuals carrying the rs225012 rSNP *DIO2*-A allele maybe at risk for sickness or disease. In fact, the rs225012 rSNP *DIO2* AA genotype [TT genotype (- strand)] frequency has been significantly associated with MR [9] in Chinese patients.

Table 1. The DIO2 SNPs that were examined in this study where the minor allele is in red. Also listed are the transcriptional factors (TF), their potential binding sites (TFBS) containing these SNPs and DNA strand orientation. TFs in red differ between the SNP alleles. Where upper case nucleotide designates the 90% conserved BS region and red is the SNP location of the alleles in the TFBS. Below the TFBS is the nucleotide occurrence (%) obtained from the Jaspar Core database. Also listed are the number (#) of binding sites in the gene for the given TF. Note: TFs can bind to more than one nucleotide sequence.

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand	
Novel Rare (-2035bp TSS)	C	BRCA1	breast cancer 1, early onset	3	ccA acc a c=67%	minus	
		FOXD3	Forkhead box D3	1	tTgTT g gt g=1%	plus	
		FOXI1	Forkhead box I1	1	tTgTT g gt g=23%	plus	
		FOXQ1	Forkhead box Q1	1	tttGTT g t g=22%	plus	
		MEIS1	Meis homeobox 1	1	accTGcCA acc aaac c=21%	minus	
		NFYA	Nuclear transcription factor Y, alpha	1	tgcatTT g TTG G ttg G=99%	plus	
		NFYB	nuclear transcription factor Y, beta	1	ctgccaa C CA A caa C=100%	minus	
		RUNX2	Runt-related transcription factor 12	1	atTTgTt G Gttg G=100%	plus	
		RUNX1	Runt-related transcription factor 1	1	ttgTt G Gttg G=92%	plus	
		SRY	Sex determining region Y	1	ccaa A CA A A c=18%	minus	
		THAP1	THAP domain containing, apoptosis associated protein 1	1	ctgCC acc c=36%	minus	
		T 0.004	ARID3A	AT rich interactive domain 3A (BRIGHT-like)	12	AT c AA a T=100%	minus
			CDX2	Caudal type homeobox 2	2	tgcca A T c AA a T=100%	minus
			FOXI1	Forkhead box I1	1	tTgTTT g att a=1%	plus

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand
rs12885300 (C/T) 5' UTR	C	FOXD3	Forkhead box D3	1	tTgTTtgatt a=13%	plus
		GFI1	Growth factor independent 1 transcription repressor	1	ccAATCaaac T=98%	minus
		HOXA9	Homeobox A9	1	ccaATcAAaCa T=100%	minus
		HOXC9	Homeobox C9	1	gccaaTcAAaaca T=99%	minus
		MEIS1	Meis homeobox 1	1	accTGcCAatcaaac t=57%	minus
		NFYB	nuclear transcription factor Y, beta	1	caacctgCCAATcaa T=100%	minus
		NKX2-5	Natural killer 3 homeobox 2	2	ttgAttg A=100%	plus
		NKX2-5 (var.2)	Natural killer 2 homeobox 5	2	tgcCaaTCaaa T=100%	minus
		Nobox	NOBOX oogenesis homeobox	1	TgATTggc A=100%	plus
		PBX1	Pre-B-cell leukemia homeobox 1	2	ctgcCAATCAaa T=94%	minus
		PBX1	Pre-B-cell leukemia homeobox 1	1	caaTCAAaCaaa T=90%	minus
		SOX17	SRY (sex determining region Y)-box 17	4	ttgATTggc A=97%	plus
		SRY	Sex determining region Y	1	tcaaACAAA t=29%	minus
		STAT3	Signal transducer and activator of transcription 3 (acute-phase response factor)	2	tTgattGGcAg a=5%	plus
		THAP1	THAP domain containing, apoptosis associated protein 1	1	ctgCCaatc t=29%	minus
		FOXP1	Forkhead box P1	1	aaaggctAAAgAaaa g=28%	plus
		PRDM1	PR domain containing 1, with ZNF domain	1	agacAatGAAAGgct	plus

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand
rs225010 (C/T)	T 0.229	SOX3	SRY (sex determining region Y)-box 3	1	g=11% cctTtcattg c=72%	minus
		TCF7L2	Transcription factor 7-like 2 (T-cell specific, HMG-box)	1	gacaaTgAAAGgct g=48%	plus
		ARID3A	AT rich interactive domain 3A (BRIGHT-like)	15	ActAAa A=100%	plus
		BATF::JUN	Basic leucine zipper transcription factor, ATF-like Jun proto-oncogene	3	tgaaaGAcTaA A=100%	plus
		FOXP1	Forkhead box P1	1	aaagactAAAgAaaa a=44%	plus
		IRF1	Interferon regulatory factor 1	1	ttagtcTTTCatTgtctctat t=12%	minus
		JUN:FOS	<i>Jun</i> proto-oncogene FBJ murine osteosarcoma viral oncogene homolog	10	TgAaagA A=90%	plus
		PAX2	Paired box gene 2	1	agtCtttc t=84%	minus
		PRDM1	PR domain containing 1, with ZNF domain	1	agacAatGAAAGact a=3%	plus
	C	SOX3	SRY (sex determining region Y)-box 3	3	cttTaGTctt T=100%	minus
		SOX6	SRY (sex determining region Y)-box 6	3	cttTaGTctt T=100%	minus
		TCF7L2	Transcription factor 7-like 2 (T-cell specific, HMG-box)	1	gacaaTgAAAGact a=25%	plus
		ELF5	E74-like factor 5	1	cttaTCCgt g=25%	plus
		ELK1	ELK1, member of ETS oncogene family	1	gtgacgGAta c=86%	minus
		GATA1	GATA binding protein 1	12	agcTTATCCcgt g=14%	plus
		GATA2	GATA binding protein 2	1	gtgagcTTATCcg g=45%	plus
		GATA4	GATA binding protein 4	1	gcTTATCcgtc	plus

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand
		JUN:FOS	<i>Jun</i> proto-oncogene FBJ murine osteosarcoma viral oncogene homolog	1	g =34% TgAcggA	minus
		KLF5	Kruppel-like factor 5 (intestinal)	1	c =67% ccgtCaCCCa	plus
		MAFB	v-maf musculoaponeurotic fibrosarcoma oncogene homolog B (avian)	1	g =1% Ggtgacgg	minus
		NFE2L1:MAFG	Nuclear factor erythroid 2-related factor 1 Transcription factor MafG	7	c =53% ggTGAc	minus
		PAX2	Paired box gene 2	1	c =76% ggtgacgg	minus
		PAX2	Paired box gene 2	1	c =55% cgtCaccc	plus
		SREBF1	Sterol regulatory element binding transcription factor 1	1	g =68% gTCAcccaaa	plus
	T	GATA1	GATA binding protein 1	3	g =28% agctTATCcat	plus
	0.415	HOXA9	Hoxa9	4	a =14% cttATccATCa	plus
		JUND (var.2)	Jun D proto-oncogene	1	A =99% atggaTaAgctcAct	minus
		KLF5	Kruppel-like factor 5 (intestinal)	1	t =25% ccatCaCCCa	plus
		MAFB	v-maf musculoaponeurotic fibrosarcoma oncogene homolog B (avian)	12	a =1% Ggtgatgg	minus
		NFATC2	Nuclear factor of activated T-cells, cytoplasmic, calcineurin-dependent 2	1	t =20% ttaTCCa	plus
		NFE2L1:MAFG	Nuclear factor erythroid 2-related factor 1 Transcription factor MafG	6	a =69% caTcAc	plus
		PAX2	Paired box gene 2	12	a =85% ggtgatgg	minus
		ZNF354C	Zinc finger protein 354C	9	t =35% atCCAt	plus
rs225011	C	CRX	Cone-rod homeobox	1	A =100% cctagGaTTAt	plus

SNP (C/T)	Allele	TFs	Protein name	# of Sites	TFBS	Strand
		ESRRA	Estrogen-related receptor alpha	1	G=100% cctAGGtCagt	minus
		ESRRB	Estrogen-related receptor beta	1	c=57% aatcctAGGTCA	minus
		GATA4	GATA binding protein 4	1	c=34% tgTTATaatcc	minus
		NKX2-5	Natural killer 3 homeobox 2	55	c=37% atAAtcc	minus
		NR5A2	Nuclear receptor subfamily 5, group A, member 2	1	c=12% taatcCtAGGtCagt	minus
		PRRX2	Paired related homeobox 2	34	c=21% gATTA	plus
		RORA_1	RAR-related orphan receptor A	2	g=7% tcctaGGTCA	minus
		RORA_2	RAR-related orphan receptor A	1	c=4% taatcTaGGTCaG	minus
		RXRA	Retinoid X receptor, alpha	1	c=1% cctAGgtCagt	minus
	T	ESRRA	Estrogen-related receptor alpha	1	c=85% tctAGGtCagt	minus
	0.415	ESRRB	Estrogen-related receptor beta	1	t=30% aattctAGGTCA	minus
		FOXL1	Forkhead box L1	2	t=47% gaattATA	plus
		GATA4	GATA binding protein 4	1	a=43% tgTTATaatc	minus
		MEF2A	Myocyte enhancer factor 2A	1	t=36% acctagAAtAataac	plus
		NKX2-5	Natural killer 3 homeobox 2	3	A=92% atAAttc	minus
		NR5A2	Nuclear receptor subfamily 5, group A, member 2	1	t=65% taattCtAGGTCAgt	minus
		PDX1	Pancreatic and duodenal homeobox 1	23	t=74% aTAATt	minus

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand	
rs225012 (A/G)	A 0.419	PRRX2	Paired related homeobox 2	68	t=61% aATTA a=88%	plus	
		RORA_1	RAR-related orphan receptor A	1	ttctaGGTCA t=60%	minus	
		RORA_2	RAR-related orphan receptor A	1	taattcTaGGTCAg t=36%	minus	
		EHF	Ets homologous factor	2	aCTTCtTc T=100%	plus	
		ELF5	E74-like factor 5	2	tactTCttc T=98%	plus	
		EN1	Engrailed homeobox 1	1	aagtagagata a=50%	minus	
		FOXC1	Forkhead box C1	4	aagaGTA a=44%	minus	
		HLTF	Helicase-like transcription factor	1	ctaCtTcttc t=58%	plus	
		HOXA5	Hoxa5	1	ctctacTt T=100%	plus	
		NKX3-2	NK3 homeobox 2	1	agaAGTaga A=100%	minus	
		PDX1	Pancreatic and duodenal homeobox 1	12	cTAcTt T=97%	plus	
		PRRX2	Paired related homeobox 2	55	aAgTA A=100%	minus	
		SPIB	Spi-B transcription factor (Spi-1/PU.1 related)	15	agaaGAA A=96%	minus	
		G	E2F6	E2F transcription factor 6	1	aaGaGGtAgag G=100%	minus
		EGR1	Early growth response 1	1	ctacCtCtctgcc C=94%	plus	
	ELF1	E74-like factor 1 (ets domain transcription factor)	1	agaagaGGtAGag G=100%	minus		
ERG	v-ets avian erythroblastosis virus E26	3	agAGGtAgaga	minus			

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand
rs225013 (G/T)	G	FOXC1	oncogene homolog Forkhead box C1	1	G=100% aagagGTA	minus
		SP1	Specificity Protein 1	3	g=25% tctCtaCCtct	plus
		SPI1	Spleen focus forming virus (SFFV) proviral integration oncogene spi1	1	C=100% cagaagaGGtAgaga	minus
		SPIB	Spi-B transcription factor (Spi-1/PU.1 related)	5	G=92% agaGGtA	minus
		ZNF263	Zinc finger protein 263	1	G=96% tggGgcagaagagtagagat	minus
		ZNF263	Zinc finger protein 263	1	g=58% gggGcAgaagagtagagata	minus
		FOXA1	Forkhead box A1	1	g=44% gtatTcTTtaCtadc	minus
		FOXC1	Forkhead box C1	10	g=19% ggctgGTA	minus
		HAND1:TCFE2 α	Heart- and neural crest derivatives-expressed protein 1: transcription factor E2A	1	G=100% aggCTGgtat	minus
		T 0.364	BATF::JUN	Basic leucine zipper transcription factor, ATF-like Jun proto-oncogene	1	g=86% gtaaaGAaTaA
	BRCA1	breast cancer 1, early onset	4	A=98% atAacag	plus	
	FOXA1	Forkhead box A1	1	a=84% ttatTcTTtaCtadc	minus	
	GFI1B	Growth factor independent 1B transcription repressor	1	t=40% gAAtaacaGcc	plus	
	HNF1A	HNF1 homeobox A	1	a=59% tGTTAtTctTtact	minus	
	HOXA5	Hoxa5	2	T=95% ctgttaTt	minus	
	MYB	v-myb myeloblastosis viral oncogene homolog	1	t=56% atAACaGcCt	plus	
					A=93%	

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand
rs225014 (C/T)	T	NFE2L1:MAFG	Nuclear factor erythroid 2-related factor 1 Transcription factor MafG	12	aaTaAc A=100%	plus
		PAX2	Paired box gene 2	2	tgttattc t=84%	minus
		SPZ1	Spermatogenic leucine zipper 1	1	agaataacagc a=50%	plus
		EGR1	Early growth response 1	1	tcacCtCCttCtgt t=14%	minus
		ELF1	E74-like factor 1 (ets domain transcription factor)	1	acagaaGGAgGtg a=31%	plus
		FOXC1	Forkhead box C1	1	cttctGTA T=100%	minus
		FOXC1	Forkhead box C1	1	ctccaGTA A=100%	plus
	C 0.421	HLTF	Helicase-like transcription factor	1	ctcCtTctgt t=25%	minus
		HOXA5	Hoxa5	1	ctgtacTg t=56%	minus
		RXR::RAR_DR 5	Retinoid X receptor: Retinoic acid receptor 5	1	aGtaCAgaaggagtgA a=1%	plus
		SPI1	Spleen focus forming virus (SFFV) proviral integration oncogene spi1	1	tacagaaGGAggtga a=42%	plus
		STAT5A::STA T5B	Signal transducer and activator of transcription 5A and transcription 5B	1	tgTctccaGtA A=90%	plus
		THAP1	THAP domain containing, apoptosis associated protein 1	1	tctCCagta a=58%	plus
		EBF1	Early B-cell factor 1	2	gtCtCcaGtGc G=100%	plus
EGR1	Early growth response 1	1	tcacCtCCttCtgc c=46%	minus		
ELF1	E74-like factor 1 (ets domain transcription factor)	1	gcagaaGGAgGtg g=41%	plus		

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand
rs6574549 (G/T)	G 0.007	HLTF	Helicase-like transcription factor	3	ctcCtTctgc c=20%	minus
		NKX3-2	NK3 homeobox 2	6	tccAGTgca g=75%	plus
		RXR::RAR_DR 5	Retinoid X receptor: Retinoic acid receptor	1	aGtgcagaaggaggtgA	plus
		SP2	Sp2 transcription factor	1	g=1% tcaCCtCCtctgca	minus
		SPIB	Spi-B transcription factor (Spi-1/PU.1 related)	7	tgcaGAA g=59%	plus
		TFAP2C	Transcription factor AP-2 gamma (activating enhancer binding protein 2 gamma)	1	catgtCtccAGtgca g=55%	plus
		ZNF354C	Zinc finger protein 354C	9	ctgCAC C=100%	minus
		FOXA1	Forkhead box A1	1	aaaaTacTtaCatat C=93%	plus
		FOXA2	Forkhead box A2	4	TacTACatatt C=90%	plus
		FOXC1	Forkhead box C1	38	aatatGTA G=100%	minus
		FOXC1	Forkhead box C1	25	tgtaaGTA g=38%	minus
		FOXD3	Forkhead box D3	1	gtaagTattttt g=55%	minus
		FOXI1	Forkhead box I1	1	acaTaTTTgtaa c=26%	plus
		FOXL1	Forkhead box L1	38	aatatgTA g=9%	minus
		FOXL1	Forkhead box L1	23	cttacATA c=43%	plus
		HLTF	Helicase-like transcription factor	4	ttaCaTattt C=100%	plus
		HLTF	Helicase-like transcription factor	4	ataCtTacat	plus

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand
		NFIL3	Nuclear factor, interleukin 3 regulated	1	c=1% aTAtGTAAgta	minus
		NKX3-1	NK3 homeobox 1	2	G=91% tTACaTA	plus
		NKX2-5	Natural killer 3 homeobox 2	3	C=95% gtAAgta	minus
		POU2F2	POU class 2 homeobox 2	1	g=6% aatacTTaCATat	plus
T		ARID3A	AT rich interactive domain 3A (BRIGHT-like)	28	C=99% ATtAa	minus
		ARID3A	AT rich interactive domain 3A (BRIGHT-like)	17	t=63% cTtAAa	plus
		FOXC1	Forkhead box C1	2	A=100% ttaaGTA	minus
		FOXD3	Forkhead box D3	1	t=19% aaaTaTTaagt	minus
		FOXI1	Forkhead box I1	1	T=98% aaaTaTTTgtaa	plus
		FOXL1	Forkhead box L1	3	a=26% cttaaATA	plus
		FOXL1	Forkhead box L1	2	a=17% ttaagTA	minus
		HLTF	Helicase-like transcription factor	1	T=30% ataCtTaat	plus
		HNF1B	HNF1 homeobox B	1	a=1% caAaTatTTAAG	minus
		HOXA5	Hoxa5	3	T=90% cttaaTa	plus
		LHX3	LIM homeobox 3	1	a=88% tacTTAAaTattt	plus
		LHX3	LIM homeobox 3	1	A=100% tatTTAAGtattt	minus
		NKX2-5	Natural killer 3 homeobox 2	4	T=100% tAAgta	minus

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand
rs225015 (A/G)	G	NKX3-1	NK3 homeobox 1	9	t=53% aTA t TTA	minus
		NKX3-2	NK3 homeobox 2	1	T =95% t t aAGTatt	minus
		BRCA1	breast cancer 1, early onset	1	t=38% ccA a ggg	minus
		EBF1	Early B-cell factor 1	1	g =14% ggacC a a G gGa	minus
		EBF1	Early B-cell factor 1	1	G =97% ttCc C ttGgtc	plus
		ELF5	E74-like factor 5	1	C =100% atctTCC c t	plus
		ESRRA	Estrogen-related receptor alpha	1	c =7% ccAA G GgaAga	minus
		HLTF	Helicase-like transcription factor	1	G =98% tcc C tTggtc	plus
		NFATC2	Nuclear factor of activated T-cells, cytoplasmic, calcineurin-dependent 2	4	C =100% tcTTCC c	plus
		NFKB1	Nuclear factor of kappa light polypeptide gene enhancer in B-cells 1	1	c =12% gGatcTT c CC t	plus
		NFKB1	Nuclear factor of kappa light polypeptide gene enhancer in B-cells 1	1	C =98% tGGatcT t CC c	plus
		PPARγ::RXRα	Peroxisome proliferator-activated receptor γ Retinoid X receptor, alpha	1	c =47% atgggacca a gggaa	minus
		REL	<i>v-rel</i> avian reticuloendotheliosis viral oncogene homolog	1	g =16% gggaagatc C	minus
		RFX5	Regulatory factor X, 5 (influences HLA class II expression)	1	g =47% tcc C ttgGtccCatt	plus
		SOX2	SRY (sex determining region Y)-box 2	1	C =95% C C tTgGTc	plus
		SOX3	SRY (sex determining region Y)-box 3	1	C =100% c c tTgGTccc	plus
SOX6	SRY (sex determining region Y)-box 6	1	c =75% c C tTgGTccc	plus		

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand
		SOX10	SRY (sex determining region Y)-box 10	11	C=90% cttggT	plus
		SPIB	Spi-B transcription factor (Spi-1/PU.1 related)	8	c=86% aagGGAA	minus
		THAP1	THAP domain containing, apoptosis associated protein 1	1	g=14% cttCCctg	plus
	A	BRCA1	breast cancer 1, early onset	2	c=68% ccAaagg	minus
	0.403	ELF1	E74-like factor 1 (ets domain transcription factor)	1	a=16% accaaaGGAAGat	minus
		ELF5	E74-like factor 5	3	a=66% atctTCct	plus
		ELK1	ELK1, member of ETS oncogene family	1	t=36% ccaaagGAag	minus
		ERG	v-ets avian erythroblastosis virus E26 oncogene homolog	3	a=7% aaAGGAAGatc	minus
		ETS1	Protein C-ets-1	1	A=96% ggatctTCcttggt	plus
		FLI1	Fli-1 proto-oncogene, ETS transcription factor	3	T=92% aaaGGAAGatc	minus
		HLTF	Helicase-like transcription factor	2	a=74% ttcCtTtggt	plus
		NFATC2	Nuclear factor of activated T-cells, cytoplasmic, calcineurin-dependent 2	8	t=58% tcTTCCt	plus
		NFKB1	Nuclear factor of kappa light polypeptide gene enhancer in B-cells 1	1	t=15% tGGatcTtCCt	plus
		RUNX1	Runt-related transcription factor 1	1	t=98% tccTttGGtcc	plus
		SOX2	SRY (sex determining region Y)-box 2	12	T=97% CCtTTGgt	plus
		SOX3	SRY (sex determining region Y)-box 3	2	t=55% cctTTGgtcc	plus
		SOX3	SRY (sex determining region Y)-box 3	2	t=87% cttTgGTccc	plus

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand
rs225017 (A/T)	T	SOX6	SRY (sex determining region Y)-box 6	2	t=25% cCtTTGgtcc	plus
		SOX9	SRY (sex determining region Y)-box 9	3	t=35% cctTtGgtc	plus
		SOX10	SRY (sex determining region Y)-box 10	14	t=5% cttTgg	plus
		SPIB	Spi-B transcription factor (Spi-1/PU.1 related)	10	t=45% aaaGGAA	minus
		SPI1	Spleen focus forming virus (SFFV) proviral integration oncogene spi1	1	a=49% gaccaaAGGAagatc	minus
		TCF7L2	Transcription factor 7-like 2 (T-cell specific, HMG-box)	1	a=71% tgggaccAAAGgaa	minus
		BATF::JUN	Basic leucine zipper transcription factor, ATF-like Jun proto-oncogene	1	A=100% gaaTGAgaaA	minus
		GATA4	GATA binding protein 4	7	a=78% tcTTATCtcat	plus
		GATA4	GATA binding protein 4	4	t=27% tTTcTCAttt	plus
		HAND1:TCFE2 α	Heart- and neural crest derivatives-expressed protein 1: transcription factor E2A	3	t=21% tgtCTGaaat	minus
		HOXA5	Hoxa5	19	a=55% ctgaaATg	minus
		JUND (var.2)	Jun D proto-oncogene	1	a=88% tgaaATGAgaaaAga	minus
		NFE2L1:MAFG	Nuclear factor erythroid 2-related factor 1 Transcription factor MafG	11	a=75% aaTGAg	minus
		PDX1	Pancreatic and duodenal homeobox 1	11	a=85% cTcAtT	plus
		SOX3	SRY (sex determining region Y)-box 3	4	T=97% cTtTTcTcat	plus
		STAT3	Signal transducer and activator of transcription 3 (acute-phase response factor)	1	t=48% tTgtctGaAAt	minus
					A=100%	

SNP	Allele	TFs	Protein name	# of Sites	TFBS	Strand
0.309	A	BATF::JUN	Basic leucine zipper transcription factor, ATF-like Jun proto-oncogene	1	gaatTGAgaaA t=7%	minus
		CEBP α	CCAAT/enhancer binding protein (C/EBP), alpha	1	tTTtCtcaAtt A=100%	plus
		CEBP β	CCAAT/enhancer binding protein (C/EBP), beta	1	ctTTtCtcaAt A=100%	plus
		GATA4	GATA binding protein 4	1	tcTTtTCtcaa a=14%	plus
		HAND1:TCFE2 α	Heart- and neural crest derivatives-expressed protein 1: transcription factor E2A	1	tgtCTGaatt t=34%	minus
		NKX2-5	Natural killer 3 homeobox 2	7	tcAAttc A=100%	plus
		NKX2-5	Natural killer 3 homeobox 2	7	tgAAttg t=59%	minus
		PRRX2	Paired related homeobox 2	68	aAtTA t=98%	minus
		SRY	Sex determining region Y	2	tttctCAAt A=96%	plus

The rs225017 rSNP *DIO2*-T allele [A (- strand) or T (+ strand)] located in the *JUND* (var.2) and *STAT3* TFBS have in humans a 75% and 100% occurrence, respectively (Table). Since these BS occurs only once in the gene, this rSNP would probably have a major impact on these TFs regulating the gene. The *HOXA5*, *NFE2L1*: *TCFE2 α* and *PDX1* TFBSs have an 88%, 85% and 97% occurrence, respectively, in humans but these BS occur more than once in the gene and consequently, the rs225017 rSNP might not have much of an impact on gene regulation by these TFs (Table). The minor rs225017 rSNP *DIO2*-A allele [T (- strand) or A (+ strand)] located in the *CEBP α* & β have a 100% occurrence in humans and are found only once in the gene. Since these BS only occur once in the gene, the SNP would probably have a major impact on these enhancer and inflammation TFs regulating the gene. The *NKX2-5*, *PRRX2* and *SRY* TFBS have in humans a 100%, 98% and 96% occurrence, respectively; however, these BS occur more than once in the gene and consequently this rSNP might not have much of an impact on *DIO2* regulation by these TFs (Table).

Similar logic can be used to evaluate the potential TFBS within the other *DIO2* rSNPs found in the Table. It should be noted that the minor -2035bp novel rSNP T allele creates ten unique potential TFBS compared to the common C allele which creates only four BS while the rs225012 rSNP *DIO2* alleles each generate eight unique potential TFBS. In fact, 57 potential TFBS are created by the minor alleles of the nine SNPs compared to 39 TFBS created by the common alleles with 51 TFBS being shared by both alleles. Since the MAF of the nine SNPs ranges from 0.004 to 0.421, the potential TFBS generated by the minor alleles should have a tremendous impact on thyroid related illnesses and other sickness in humans. As an example, the *POU2F2* (*POU* class 2 homeobox 2) TFBS is only created by the minor rare allele of rs6574549 and occurs only once in the gene which is important because it's a TF that binds in immunoglobulin gene promoters (supplement). This rSNP has been associated with fasting insulin, insulin action and energy expenditure in Pima Indians [11].

Human diseases or conditions can be associated with rSNPs of the *DIO2* gene as illustrated above. What a change in the rSNP alleles can do, is to alter the DNA landscape around the SNP for potential TFs to attach and regulate a gene. As an example, the potential TFBS

associated with the novel -2035bp common rSNP *DIO2*-C allele from Table are illustrated in Fig. 2 as well as the rs225015 rSNP *DIO2*-G allele illustrated in Fig. 1. As can be seen in Table, these potential TFBS change when an individual carries the minor allele. The importance of this can be illustrated with the *BRCA1* TFBS where the common allele has this function and the minor allele does not. The *BRCA1* TF plays a role in maintaining genomic stability and also acts as a tumor suppressor. Another example would be the *PPARG::RXRA* TFBS where the common allele has this function while the minor allele does not. This TF has been implicated in the pathology of numerous diseases including obesity, diabetes, atherosclerosis and cancer.

5. CONCLUSION

SNPs that alter the TFBS are not only found in the promoter regions but in the introns, exons and the UTRs of a gene (Table). The nucleus of the cell is where epigenetic alterations occur and TFs operate to convert chromosomes into single stranded DNA for mRNA transcription while it is the cytoplasm where mRNA is processed by separating exons and introns for protein translation. Consequently, it doesn't matter where TFs bind the DNA in the nucleus because it is only there that TFs function. The SNPs outlined in this report should be considered as rSNPs since they change the DNA landscape for TF binding and have been associated with disease. In this report, examples have been described to illustrate that a change in rSNP alleles in the *DIO2* gene can provide different TFBS which in turn are also associated with disease in humans. The potential alterations in TFBS obtained by computational analyses need to be verified by future protein/DNA electrophoretic mobility gel shift assays and gene expression studies.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Author has declared that no competing interests exist.

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